

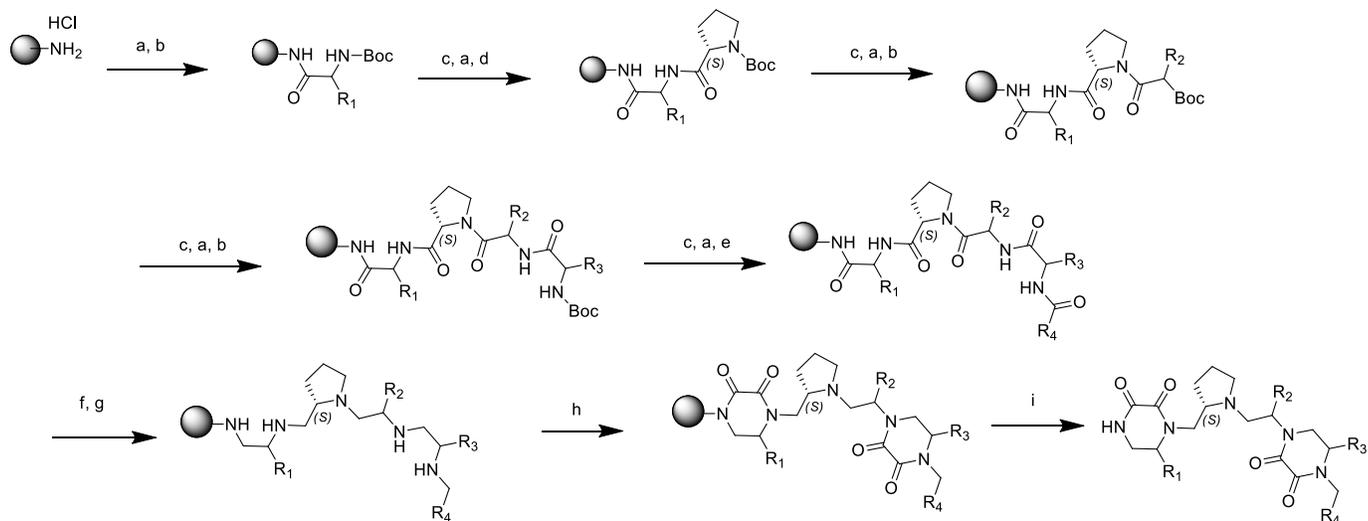
Supplemental Material

A Novel Probe for Spliceosomal Proteins that Induces Autophagy and Death of Melanoma Cells Reveals New Targets for Melanoma Drug Discovery

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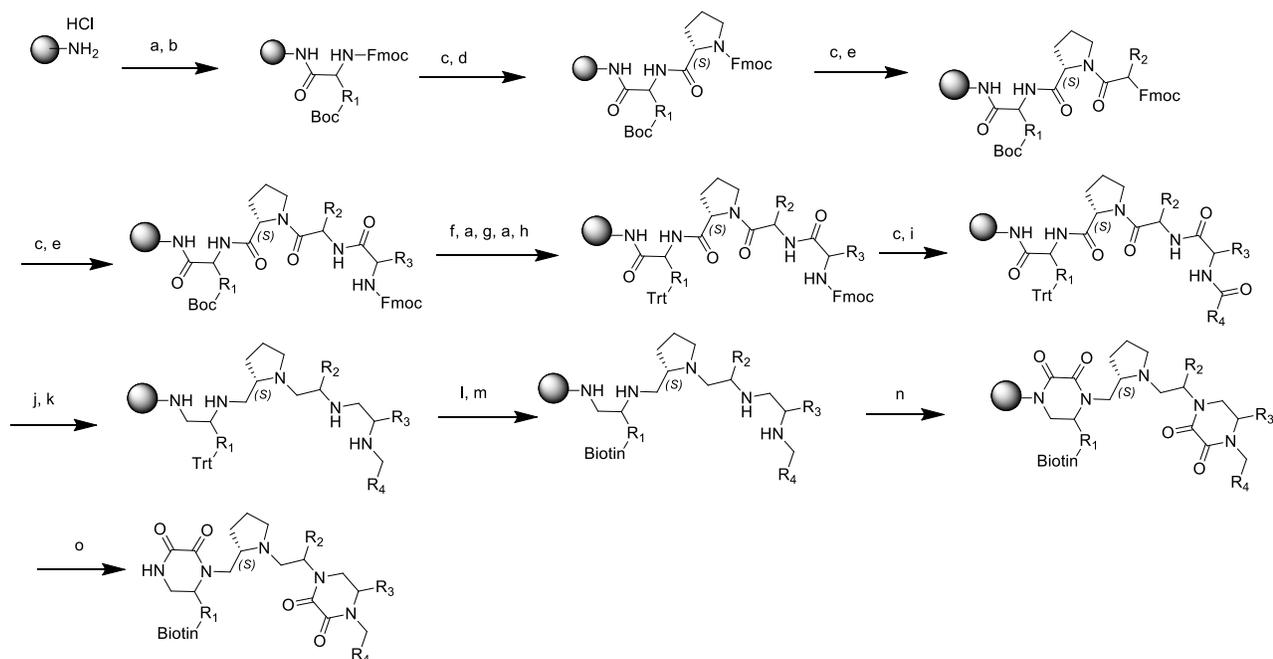
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Compound purification and characterization.



Supplementary Figure 1A. Synthesis procedure for (S)-1((R)-1((S)-2-(((S)-6-benzyl-2,3-dioxopiperazin-1-yl)methyl)pyrrolidin-1-yl)-3-cyclohexylpropan-2-yl)-5-(4-hydroxybenzyl)-4-phenethylpiperazine-2,3-dione:

(a) 5% DIEA/95% DCM, 3x 2min; (b) Boc-AA (6eq), DIC (6eq), HOBT (6eq), DMF, 2hr; (c) 55% TFA/45% DCM, 30min; (d) Boc-L-Pro-OH (6eq), DIC (6eq), HOBT (6eq), DMF (6eq), 2hr; (e) RCOOH (10eq), DIC (10eq), HOBT (10eq), DMF, 2hr; (f) BH₃/THF (40eq), 65°C, 96hr; (g) Piperidine, 65°C, 18hr; (h) (COIm)₂ (10eq), Anhydrous Atmosphere, 18hr; (i) HF/Anisole, 0°C, 1.5hr



Supplementary Figure 1B. Synthesis procedure for Biotin tagged pyrrolidine-bis-diketopiperazines used in present studies: (a) 5% DIEA/95% DCM, 3x 2min; (b) Fmoc-AA(Boc) (6eq), DIC (6eq), HOBT (6eq), DMF, 2hr; (c) 20% Piperidine/80% DMF, 2x 30min; (d) Fmoc-L-Pro-OH (6eq), DIC (6eq), HOBT (6eq), DMF (6eq), 2hr; (e) Fmoc-AA (6eq), DIC (6eq), HOBT (6eq), DMF, 2hr; (f) 55% TFA/45% DCM, 30min; (g) Trt-Cl (5eq), DIEA (10eq), 10% DMF/90% DCM, 2hr; (h) Trt-Cl (5eq), DIEA (10eq), 10% DMF/90% DCM, 2hr; (i) RCOOH (10eq), DIC (10eq), HOBT (10eq), DMF, 2hr; (j) BH₃/THF (40eq), 65°C, 96hr; (k) Piperidine, 65°C, 18hr; (l) 2% TFA/5% TRIS, 95% DCM, 3x 2min; (m) Biotin (10eq), DIC (10eq), DMF, 2hr; (n) (COIm)₂ (10eq), Anhydrous Atmosphere, 18hr; (o) HF/Anisole, 0°C, 1.5hr

(S)-1((R)-1((S)-2-(((S)-6-benzyl-2,3-dioxopiperazin-1-yl)methyl)pyrrolidin-1-yl)-3-cyclohexylpropan-2-yl)-5-(4-hydroxybenzyl)-4-phenethylpiperazine-2,3-dione. Using General Scheme (scheme 1) for the synthesis of bis-cyclic diketopiperazines compounds 2155-14 and 2529-1 were synthesized using the following reagents: (2 g) MBHA resin starting material, Boc-L-Phenylalanine -OH (R₁), Boc-D-Cyclohexylalanine-OH (R₂), Boc-L-Tyrosine(2-Br-Z)-OH (R₃), and Phenylacetic Acid (R₄). The final crude product was purified using HPLC as described above, with a gradient of (B) 0/5, 2/5, 4/20, 40/55. **Isolated Mass** 504.2mg, **%yield** 28.62%.

¹H NMR (400 MHz, DMSO-*d*₆) δ ppm 0.80 - 1.05 (m, 2 H) 1.08 - 1.20 (m, 3 H) 1.24 (br. s., 2 H) 1.50 - 1.71 (m, 6 H) 1.78 - 1.96 (m, 2 H) 1.97 - 2.13 (m, 2 H) 2.55 - 2.65 (m, 2 H) 2.67 - 2.88 (m, 6 H) 2.90 - 3.04 (m, 3 H) 3.11 (d, *J*=12.96 Hz, 1 H) 3.19 - 3.32 (m, 2 H) 3.37 (br. s., 1 H) 3.52 (dd, *J*=13.08, 3.30 Hz, 2 H) 3.65 (d, *J*=10.03 Hz, 2 H) 3.70 - 3.94 (m, 2 H) 6.73 (m, *J*=8.07 Hz, 2 H) 7.01 (m, *J*=8.07 Hz, 2 H) 7.15 - 7.31 (m, 9 H) 8.50 (d, *J*=5.01 Hz, 1 H) **m/z** calcd C₄₄H₅₅N₅O₅ [M+H]⁺ 734.42, found 734.15 (MALDI), 734.15 (MS ESI) **Purity** LC-MS: 99.0% (254 nm, peak height).

N-(4-((S)-1-(((S)-1-((R)-3-cyclohexyl-2-((S)-5-(4-hydroxybenzyl)-2,3-dioxo-4-phenethylpiperazin-1-yl)propyl)pyrrolidin-2-yl)methyl)-5,6-dioxopiperazin-2-yl)butyl)-5-((3aS,4S,6aR)-2-oxohexahydro-1H-thieno[3,4-d]imidazol-4-yl)pentanamide. Using Scheme 3 for the synthesis of biotin tagged pyrrolidine-bis-diketopiperazines compound 2529-3 was synthesized using the following reagents: (100 mg) MBHA resin starting material, Fmoc-L-Lysine(Boc) -OH (R₁), Fmoc-D-Cyclohexylalanine-OH (R₂), Fmoc-L-Tyrosine(2-Br-Z)-OH (R₃), and Phenylacetic Acid (R₄). The final crude product was purified using HPLC as described above, with a gradient of (B) 0/5, 2/5, 4/20, 40/55. **Isolated Mass** 4.9mg **% yield** 4.33%.

¹H NMR (400 MHz, DMSO-*d*₆) δ ppm 9.63 (br. s., 1H) 9.35 (br. s., 1H) 8.56 (br. s., 1 H) 7.81 (br. s., 1 H) 7.29 (br. s., 1 H) 7.24 (br. s., 2 H) 7.01 (br. s., 1 H) 6.72 (br. s., 1 H) 6.40 (d, J=15.89 Hz, 1 H) 4.90 (br. s., 1 H) 4.31 (br. s., 1 H) 3.93 - 4.17 (m, 1 H) 3.63 - 3.87 (m, 4 H) 3.60 (br. s., 1 H) 3.53 (br. s., 2 H) 3.44 (br. s., 14 H) 3.31 (br. s., 2 H) 3.20 (br. s., 2 H) 3.09 (br. s., 1 H) 3.04 (br. s., 1 H) 2.83 (d, J=11.49 Hz, 3 H) 2.74 (br. s., 1 H) 2.59 (d, J=12.72 Hz, 4 H) 2.28 (br. s., 1 H) 2.04 (br. s., 3 H) 1.89 (br. s., 1 H) 1.81 (d, J=9.41 Hz, 1 H) 1.69 (br. s., 2 H) 1.63 (br. s., 3 H) 1.48 (br. s., 2 H) 1.42 (br. s., 2 H) 1.29 (br. s., 4 H) 1.19 (br. s., 2 H) 0.96 (br. s., 2 H) **m/z** calcd C₅₁H₇₂N₈O₇S [M+H]⁺ 941.53, found 471.15 (MS ESI) **Purity** LC-MS: 99.0% (254 nm, peak height).

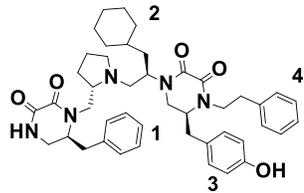
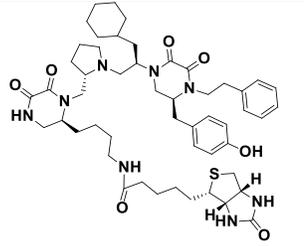
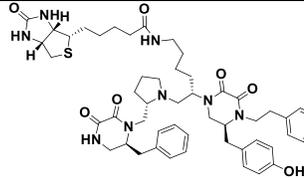
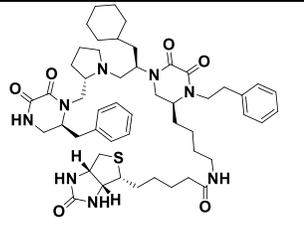
N-((S)-6-((S)-2-(((S)-6-benzyl-2,3-dioxopiperazin-1-yl)methyl)pyrrolidin-1-yl)-5-((S)-5-(4-hydroxybenzyl)-2,3-dioxo-4-phenethylpiperazin-1-yl)hexyl)-5-((3aS,4S,6aR)-2-oxohexahydro-1H-thieno[3,4-d]imidazol-4-yl)pentanamide. Using Scheme 3 for the synthesis of Biotin tagged pyrrolidine-bis-diketopiperazines compound 2529-5 was synthesized using the following reagents: (100mg) MBHA resin starting material, Fmoc-L-Phenylalanine -OH (R₁), Fmoc-L-Lysine(Boc) -OH (R₂), Fmoc-L-Tyrosine(2-Br-Z)-OH (R₃), and Phenylacetic Acid (R₄). The final crude product was purified using HPLC as described above, with a gradient of (B) 0/5, 2/5, 4/20, 40/55. **Isolated Mass** 6.4 mg **% yield** 5.70%.

¹H NMR (400 MHz, DMSO-*d*₆) δ ppm 9.53 (br. s., 1 H) 9.35 (br. s., 1 H) 8.62 (br. s., 1 H) 7.82 (br. s., 1 H) 7.25 - 7.35 (m, 6 H) 7.22 (br. s., 1 H) 7.01 (br. s., 2 H) 6.72 (br. s., 1 H) 6.30 - 6.46 (m, 1 H) 4.76 (br. s., 1 H) 4.30 (br. s., 1 H) 4.12 (br. s., 1 H) 4.02 (d, J=12.47 Hz, 1 H) 3.78 (br. s., 2 H) 3.56 - 3.69 (m, 4 H) 3.51 (br. s., 10 H) 3.33 (br. s., 2 H) 3.16 (d, J=11.62 Hz, 2 H) 2.92 - 3.11 (m, 4 H) 2.76 - 2.92 (m, 4 H) 2.70 (br. s., 1 H) 2.57 (d, J=12.10 Hz, 1 H) 2.26 (br. s., 1 H) 2.02 (br. s., 3 H) 1.84 (br. s., 1 H) 1.59 (br. s., 1 H) 1.43 (br. s., 5 H) 1.29 (br. s., 2 H) **m/z** calcd C₅₁H₆₆N₈O₇S [M+H]⁺ 935.48, found 468.45 (MS ESI) **Purity** LC-MS: 99.0% (254 nm, peak height).

N-(4-((S)-4-((R)-1-((S)-2-(((S)-6-benzyl-2,3-dioxopiperazin-1-yl)methyl)pyrrolidin-1-yl)-3-cyclohexylpropan-2-yl)-5,6-dioxo-1-phenethylpiperazin-2-yl)butyl)-5-((3aR,4R,6aS)-2-oxohexahydro-1H-thieno[3,4-d]imidazol-4-yl)pentanamide. Using Scheme 3 for the synthesis of Biotin tagged pyrrolidine-bis-diketopiperazines compound 2529-7 was synthesized using the following reagents: (100mg) MBHA resin starting material, Fmoc-L-Phenylalanine -OH (R₁), Fmoc-D-Cyclohexylalanine-OH (R₂), Fmoc-L-Lysine(Boc) -OH (R₃), and Phenylacetic Acid (R₄). The final crude product was purified using HPLC as described above, with a gradient of (B) 0/5, 2/5, 4/20, 40/55. **Isolated Mass** 16.1mg **%yield** 14.50%.

¹H NMR (400 MHz, DMSO-*d*₆) δ ppm 9.39 (br. s., 1 H) 8.65 (br. s., 1 H) 7.78 (br. s., 1 H) 7.33 (br. s., 5 H) 7.28 (br. s., 3 H) 6.29 - 6.47 (m, 1 H) 4.86 (br. s., 1 H) 4.21 - 4.36 (m, 1 H) 4.12 (br. s., 1 H) 4.03 (d, J=12.72 Hz, 2 H) 3.79 (br. s., 4 H) 3.59 - 3.75 (m, 7 H) 3.52 (d, J=18.58 Hz, 3 H) 3.25 - 3.43 (m, 2 H) 3.18 (br. s., 2 H) 2.97 - 3.10 (m, 4 H) 2.92 (br. s., 2 H) 2.70 - 2.87 (m, 1 H) 2.57 (d, J=11.62 Hz, 1 H) 2.29 (br. s., 1 H) 2.04 (br. s., 2 H) 1.99 (br. s., 1 H) 1.85 (br. s., 2 H) 1.65 (br. s., 2 H) 1.60 (br. s., 3 H) 1.48 (br. s., 2 H) 1.33 - 1.44 (m, 3 H) 1.29 (br. s., 4 H) 1.13 (br. s., 3 H) 0.72 - 0.96 (m, 2 H) **m/z** calcd C₅₁H₇₂N₈O₆S [M+H]⁺ 925.53, found 463.20 (MS ESI) **Purity** LC-MS: 95.0% (254 nm, peak height).

Supplemental Table 1. Viability testing of biotinylated analogs of compound 2155-14 with WM266-4 cells. All units are IC₅₀, μM (n=3). Numbers in 2155-14 structure indicate positions of substitutions of basic scaffold.

ID	Structure	R1	R2	R3	R4	IC ₅₀ , μM
2155-14/2529-1						4.0/1.6
2529-3		Biotin				>100
2529-5			Biotin			>100
2529-7				Biotin		3.3

Supplementary Figure 2. Comparison of sequence coverage of bands 3 and 4 from pulldown experiment. Yellow = identified peptides, green = modified amino acids. Top = coverage of band 4, bottom = coverage of band 3. Please note that there is a difference in coverage of amino acids 3-14 between band 3 and 4 suggesting that band 3 is hnRNP B1 and band 4 is hnRNP A2.

P22626 (100%), 37,430.3 Da

Heterogeneous nuclear ribonucleoproteins A2/B1 n=12 Tax=Boreoeutheria RepID=ROA2_HUMAN

22 exclusive unique peptides, 34 exclusive unique spectra, 89 total spectra, 189/353 amino acids (54% coverage)

MEK T L E T V P L	ER K K R E K E Q F	R K L F I G G L S F	ET T E E S L R N Y	Y E Q W G K L T D C
V V M R D P A S K R	S R G F G F V T F S	S M A E V D A A M A	A R P H S I D G R V	V E P K R A V A R E
E S G K P G A H V T	V K K L F V G G I K	E D T E E H H L R D	Y F E E Y G K I D T	I E I I T D R Q S G
K K R G F G F V T F	D D H D P V D K I V	L Q K Y H T I N G H	N A E V R K A L S R	Q E M Q E V Q S S R
S G R G G N F G F G	D S R G G G G N F G	P G P G S N F R G G	S D G Y G S G R G F	G D G Y N G Y G G G
P G G G N F G G S P	G Y G G G R G G Y G	G G G P G Y G N Q G	G G Y G G G Y D N Y	G G G N Y G S G N Y
N D F G N Y N Q Q P	S N Y G P M K S G N	F G G S R N M G G P	Y G G G N Y G P G G	S G G S G G Y G G R
S R Y				

P22626 (100%), 37,430.3 Da

Heterogeneous nuclear ribonucleoproteins A2/B1 n=12 Tax=Boreoeutheria RepID=ROA2_HUMAN

23 exclusive unique peptides, 30 exclusive unique spectra, 58 total spectra, 219/353 amino acids (62% coverage)

MEK T L E T V P L	ER K K R E K E Q F	R K L F I G G L S F	ET T E E S L R N Y	Y E Q W G K L T D C
V V M R D P A S K R	S R G F G F V T F S	S M A E V D A A M A	A R P H S I D G R V	V E P K R A V A R E
E S G K P G A H V T	V K K L F V G G I K	E D T E E H H L R D	Y F E E Y G K I D T	I E I I T D R Q S G
K K R G F G F V T F	D D H D P V D K I V	L Q K Y H T I N G H	N A E V R K A L S R	Q E M Q E V Q S S R
S G R G G N F G F G	D S R G G G G N F G	P G P G S N F R G G	S D G Y G S G R G F	G D G Y N G Y G G G
P G G G N F G G S P	G Y G G G R G G Y G	G G G P G Y G N Q G	G G Y G G G Y D N Y	G G G N Y G S G N Y
N D F G N Y N Q Q P	S N Y G P M K S G N	F G G S R N M G G P	Y G G G N Y G P G G	S G G S G G Y G G R
S R Y				

Supplementary Figure 3. Optimization of DDX1, hnRNP H2, and hnRNP A2/B1 siRNA knock-down conditions. Encircled concentrations of respective siRNAs were chosen for downstream experiments. C1 = scrambled siRNA control at 100nM.

