Supplementary Material

Endothelial-Specific Overexpression of Histone Deacetylase 2 Protects Mice against Endothelial Dysfunction and Atherosclerosis

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Hindlll digestion of HDAC2 clones. The Hindll digestion of HDAC2 clones. The preserving the amino acid coded by that codon. (**B**) HDAC activity from HEK 293 cells expressing either wild-type HDAC2 or mutant HDAC2 lacking the Hindlll site was determined with the HDAC Fluorometric Activity Assay Kit (Cayman). (**C**) Sall restriction digestion of human HDAC2 cDNA that was subcloned into pSPTg.T2FpAXK (30 μ g) utilizing unique Hindlll and Notl restriction sites. (**D**) Primers specific to human HDAC2 were used for PCR analysis with genomic DNA isolated from the tails of the following mice as templates: for (A) C57BL/6 (lane 2), non-transgenic littermates (lane 3), and an HDAC2 transgenic founder (lane 4); a transgenic DNA construct that was injected into the murine ooytes was used as positive control template (lane 1).



Supplementary Figure S2. HDAC2 protects endothelial-dependent vascular relaxation in atherogenic ApoE^{-/-} **mice.** Dose-response (D/R) to (A) acetylcholine (Ach), **p<0.005 vs. C57BL6/Ad.vector. (B) sodium nitroprusside (SNP) in aortic rings isolated from ApoE^{-/-} mice fed with a high-fat diet (HFD) and administered three injections of either adenoviral HDAC2 or Ad-vector (control). (C) HDAC2 expression in aortas isolated from HDAC2 adenovirus-injected mice fed a HFD regimen for 10 weeks.



expression in myeloid cells. (A) Snap-frozen, isolated endothelium-intact (E+, control) and endothelium-denuded (E-) aortas from HDAC2-Tg mice were subjected to western blotting using anti-HDAC2, anti-eNOS, and anti-GAPDH (loading control) antibodies. (B) Bone marrow was harvested from femurs and subjected to western blotting using anti-HDAC2 and anti-GAPDH antibodies. IB, immunoblot; WT, wild-type.



After 48 hours, dose-response effects of **(B)** acetylcholine (*p<0.05 vs. WT+OxLDL) and **C)** sodium nitroprusside on vascular relaxation were determined by wire myography. **(D)** "*En face*" detection of NO production in aortic intima of WT and HDAC2-Tg mice was measured by DAF fluorescence and presented as relative fluorescence unit (RFU). *p<0.05 vs. WT+HDAC2-Tg.



Supplementary Figure S5. Mouse model of atherosclerosis with PCSK9 mutant. (A) Atherosclerosis was induced in C57BL/6 mice via one intraperitoneal injection of adenoassociated virus encoding murine PCSK9 D377Y gain-of-function cDNA controlled by a liverspecific promoter. Liver lysates were immunoblotted for PCSK9, low-density lipoprotein receptor (LDL-r), and β -actin. (B) Densitometric analyses of western blot shown in A (*p<0.05 vs. PBS). (C) Non-HDL cholesterol was determined in phosphate-buffered saline- and PCSK9 AAVinjected mice (*p<0.05 vs. PBS [Con]).

