

Review

Pathophysiological Role of K_{2p} Channels in Human Diseases

Li-Ming Lee^a Thomas Müntefering^b Thomas Budde^c Sven G. Meuth^b
Tobias Ruck^b

^aDepartment of Neurology with Institute of Translational Neurology, University Hospital Münster, Münster, Germany, ^bDepartment of Neurology, University Hospital Düsseldorf, Düsseldorf, Germany, ^cInstitute of Physiology I, Westfälische Wilhelms-Universität, Münster, Germany

Key Words

K_{2p} channels • Pathophysiological mechanisms • Oncology • CNS disorders • Autoimmune diseases • Cardiovascular diseases • Hematologic diseases • Type 2 diabetes • Urinary and GI disorders

Abstract

The family of two-pore domain potassium (K_{2p}) channels is critically involved in central cellular functions such as ion homeostasis, cell development, and excitability. K_{2p} channels are widely expressed in different human cell types and organs. It is therefore not surprising that aberrant expression and function of K_{2p} channels are related to a spectrum of human diseases, including cancer, autoimmune, CNS, cardiovascular, and urinary tract disorders. Despite homologies in structure, expression, and stimulus, the functional diversity of K_{2p} channels leads to heterogeneous influences on human diseases. The role of individual K_{2p} channels in different disorders depends on expression patterns and modulation in cellular functions. However, an imbalance of potassium homeostasis and action potentials contributes to most disease pathologies. In this review, we provide an overview of current knowledge on the role of K_{2p} channels in human diseases. We look at altered channel expression and function, the potential underlying molecular mechanisms, and prospective research directions in the field of K_{2p} channels.

© 2021 The Author(s). Published by
Cell Physiol Biochem Press GmbH&Co. KG

Introduction

Two-pore domain potassium (K_{2p}) channels have been identified and broadly characterized in the last decades. While the concept of background potassium currents was discovered by Bernstein in 1902, the first K_{2p} channel in drosophila, c.elegans, and mammals was identified in 1996 by the groups of Goldstein and Lesage [1, 2]. Being initially recognized as a mere background leak channel, the relevance of K_{2p} channels to human disease was for a long time unknown. Today, K_{2p} channels are well known not only

for contributing to potassium leak currents but also for maintaining the resting membrane potential and modulating diverse physiological functions in mammalian cells. There are fifteen members in the K_{2p} channel family, divided into six subgroups (THIK, TASK, TRESK, TWIK, TALK, and TREK) according to their distinct primary structures, physiological properties, and biological functions. All K_{2p} channel members share common structural features, with two pore-forming loops and four transmembrane domains (4TMD) with intracellular amino- and carboxyl-termini. Also, they function as homo- or heterodimers instead of tetramers as in other potassium channels [3].

The opening of K_{2p} channels is mainly voltage-independent but highly regulated by stimuli such as temperature, pH, mechanical stretch, lipids, and anesthetics [4, 5]. K_{2p} channels are insensitive to typical potassium channel blockers [6]. They are broadly expressed throughout the human body with specific expression profiles among the subgroups (Table 1). For example, almost all K_{2p} channels are highly expressed in the central nervous system (CNS), while only a few show prominent expression in liver, gallbladder, and lung. Among the different subgroups, TASK and TRESK subfamilies are highly expressed in endocrine and reproductive systems, whereas TWIK and TALK subfamilies are mainly observed in cardiac and gastrointestinal systems, lymphoid organs, and pancreas. The K_{2p} channel expression profile is an essential indicator of physiological function. On a cellular level, K_{2p} channel functions in excitable cells have been studied intensely, and lately, their influence on non-excitabile cells became evident [7]. In excitable cells, K_{2p} channels modulate cellular activity and muscle tone through stabilizing the action potential in neuronal and cardiac systems and contributing to general physiological functions such as thermosensation, nociception, and muscle contraction/relaxation [8-12]. In non-excitabile cells, their prominent expression in the pancreas, immune cells, kidney, and cancer cells associate them with a wide range of pathological conditions, including type 2 diabetes, multiple sclerosis (MS), cancer, and renal disorders [13-16]. In these diseases, the aberrant function of K_{2p} channels influences insulin secretion, T cell activation/proliferation, blood-brain barrier function as well as potassium re-absorption and homeostasis. Accordingly, TASK1^{-/-} mice revealed an attenuated disease course in experimental autoimmune encephalomyelitis (EAE), the animal model of multiple sclerosis, by reducing the activation and effector functions of T lymphocytes [17-19].

Table 1. K_{2p} channel RNA expression profile in human tissues. The table demonstrates the RNA expression levels of fifteen K_{2p} channels in human tissues following the online database The Human Protein Atlas (<https://www.proteinatlas.org/>). In general, the expression profile varies depending on the individual K_{2p} channel; some are expressed heterogeneously, whereas others are expressed in specific tissues. Channel names are based on the HUGO nomenclature. The indication of channel expression levels in different human tissues is categorized as abundant (+++), mild (++) , and little (+). CNS: central nervous system, GI: gastrointestinal system

Channel HUGO name	CNS	Endocrine	Reproductive system	Liver	Cardiac	Kidney/ urinary bladder	GI	Lymphoid and blood
KCNK13	++	+++	+++			+		+
KCNK12	+++	+		++				+
KCNK9								
KCNK15		+	+++	++				
KCNK3	+	+++	+					
KCNK18	+		+					
KCNK1	++		+				+	
KCNK6		++	+++		++	++	+	+++
KCNK7							++	+
KCNK5		+	+	++		++	+++	
KCNK17		+	+		+			+++
KCNK16		+++					++	+
KCNK2	+	+++						
KCNK10	+++						+++	+++
KCNK4	+++							

A TASK3 channel blocker displayed its efficacy in inhibiting tumor formation by affecting cell cycle and proliferation in tumors [20, 21]. While some K_{2p} channels have been thoroughly investigated in various pathological conditions, others remain poorly characterized.

We here provide a comprehensive overview of current knowledge about the role of K_{2p} channels in human disease (Table 2) as well as novel ideas for mechanistic studies to further unravel the pathophysiological interrelations and identify potential therapeutic strategies (Table 3).

K_{2p} channels in oncology

Background

Cancer is a phenomenon of uncontrolled cell growth and metastatic dissemination. It is caused by both genetic and environmental factors [22, 23] that lead to distinct cellular alterations such as abnormal cellular metabolism, epigenetic changes, and increased angiogenesis [24, 25]. Oncogenes have been suggested to activate potassium channels and promote tumorigenesis [26, 27]. Thirteen out of fifteen K_{2p} channel was found to associate with carcinogenesis, each K_{2p} channel plays a distinct role in tumor development, depending on its biophysical characteristics, sensitivity to different stimuli (pH, hypoxia, reactive oxygen species, stretch, calcium and glucose levels), and expression patterns.

Human data

Summary. In 2013, a systemic screening of the mRNA expression of fifteen K_{2p} channels was conducted in 20 types of cancer by using OncoPrint, an online cancer microarray database, together with a meta-analysis. Despite heterogeneous expression of K_{2p} channels among different cancers, overexpression of *KCNK1* and *KCNK15* was observed in most cancers (bladder, breast, cervical, lung, pancreatic, head and neck cancer, and leukemia), whereas *KCNK3* and *KCNK10* were downregulated in breast, lung, pancreatic cancer, and sarcoma [13, 28].

Breast cancer. Ten out of fifteen K_{2p} channels showed altered expression in breast cancer. Among those ten genes, eight were upregulated (*KCNK1*, *KCNK3*, *KCNK5*, *KCNK6*, *KCNK9*, *KCNK13*, *KCNK15*, and *KCNK17*), whereas *KCNK2* and *KCNK10* were downregulated. Upregulation of *KCNK5*, *KCNK9*, and *KCNK2* was associated with triple-negative type breast cancer (TNBC), which is characterized by poor prognosis and limited treatment options [28, 29]. Therefore, targeting K_{2p} channels might be a potential therapeutic strategy. Functionally, K_{2p} channels modulate breast cancer development in cell proliferation, metastasis, and apoptosis [30, 31]. *KCNK2* was upregulated in the MDA-MB-231BO human metastasis cancer cell line and is highly related to the metastasis towards bone by modulating bone sialoprotein (BSP) and its downstream factor $\alpha\beta 3$ integrin [32].

Leukemia. Overexpression of *KCNK3*, *KCNK10*, and *KCNK12* and downregulation of *KCNK6* was found in patients with leukemia, resistant hematopoietic cancers, and acute myeloid leukemia (AML) [13, 28]. *KCNK15* was linked with acute lymphoid leukemia by altering DNA methylation in peripheral blood mononuclear cells (PBMCs) from patients [33]. On the other hand, TRESK was detected in patients with acute lymphoblast leukemia and lymphoma, indicating a regulatory role in lymphocyte proliferation and tumorigenesis [34].

Melanoma. TASK3 intracellular positivity was found in human melanoma tissue and three primary and metastatic human melanoma cell lines (WM35, HT199, and HT168-M1) [35], while *KCNK5* was downregulated in melanoma patient samples [13]. *KCNK7* was suggested as a disease biomarker and molecular target in melanoma patient specimens from a study concerned with the in silico identification and experimental validation [36].

Table 2. Role of K_{2p} channels in different pathological conditions The table summarizes all the findings on K_{2p} channels relevant to human diseases, including altered channel expression, channel mutation, and genetic variants associated with different pathological conditions. The channel name is based on the IUPHAR nomenclature. TNBC: triple-negative breast cancer, ALL: acute lymphoblastic leukemia, HCC: hepatocellular carcinoma, MS: multiple sclerosis, SNP: single nucleotide polymorphism, SCD: sickle cell disease, PAH: pulmonary artery hypertension, IVF: idiopathic ventricular fibrillation, LQTS: long QT syndrome, FHEIG: facial dysmorphism, hypertrichosis, epilepsy, intellectual/developmental delay, and gingival overgrowth syndrome, BBMRs: Birk Barel mental retardation syndrome, SSRIs: selective serotonin reuptake inhibitors, MDD: major depressive disorder, IGE: Idiopathic generalized epilepsies, T2D: type 2 diabetes, MODY: maturity-onset diabetes of the young, BEN: Balkan endemic nephropathy, DO: detrusor overactivity, LUTS: lower urinary tract symptoms, HSCR: Hirschsprung disease. CNS: central nervous system, GI: gastrointestinal system

Disease type	Channel name	Remarks and effect	References	
Oncology	THIK1 (K _{2p} 13.1)	Upregulation in breast cancer		
	TASK4 (K _{2p} 12.1)	Upregulation in resistant hematopoietic tumors and leukemia Downregulation in astrocytoma and glioblastoma	[13, 28]	
	TASK3 (K _{2p} 9.1)	Upregulation in breast cancer, TNBC, and melanoma	[29, 35]	
	TASK5 (K _{2p} 15.1)	Association of DNA demethylation in leukemia, breast, gastric, and pancreatic cancers	[33, 39, 45]	
	TASK1 (K _{2p} 3.1)	Upregulation in leukemia, lymphedema, breast, and kidney cancers	[13]	
	TWIK1 (K _{2p} 1.1)	Upregulation in most cancers Downregulation in melanoma, prostate cancers, and sarcoma	[13]	
	TWIK2 (K _{2p} 6.1)	Downregulation in AML, colorectal, esophageal cancers, leukemia, and melanoma Upregulation in breast and ovary cancer		
	KCNK7 (K _{2p} 7.1)	Downregulation in cervical cancer and potential marker for melanoma	[43]	
	TRESK (K _{2p} 18.1)	Upregulation in acute lymphoblast leukemia and lymphoma	[28, 34]	
	TASK2 (K _{2p} 5.1)	Downregulation in melanoma Upregulation in breast, pancreatic cancer, and HCC	[13, 29, 37]	
	TASK4 (K _{2p} 12.1)	Upregulation in breast cancer Downregulation in HCC and CNS cancers	[13, 28, 37]	
	TREK1 (K _{2p} 2.1)	Downregulation in breast cancer and HCC Upregulation in prostate cancer	[28, 37, 43]	
	TREK2 (K _{2p} 10.1)	Downregulation in most cancers and mediate breast cancer via estrogen receptor	[13]	
	Autoimmune disease	TASK1 (K _{2p} 3.1)	Downregulation in MS patient samples	[19]
		TASK2 (K _{2p} 5.1)	Upregulation in MS and Rheumatoid arthritis patient samples	[18, 54]
TASK3 (K _{2p} 9.1)		Downregulation in MS and UC patient samples	[19, 53]	
Hematologic disease	TWIK2 (K _{2p} 6.1)	KCNK6 is associated with the SNPs in SCD patients	[69]	
Cardiovascular disorders	TASK1 (K _{2p} 3.1)	Common target for class III antiarrhythmic drugs and A293 Missense mutation is associated with PAH	[90, 93, 95]	
	TASK2 (K _{2p} 5.1)	Risk genes for coronary arterial disorders and myocardial infarction patients	[89]	
	TALK2 (K _{2p} 17.1)	Mutation with loss of channel functions in IVF, heart failure, and LQTS patients Genetic variants in cerebral hemorrhage and ischemic stroke	[84, 85, 100, 103]	
	TREK1 (K _{2p} 2.1)	Downregulation in atrial fibrillation, and tachycardia patients	[97, 100]	
CNS disease	TAAK (K _{2p} 4.1)	Missense mutations contribute to neurodevelopmental disease, FHEIG	[117]	
	TASK3 (K _{2p} 9.1)	Genetic mutation and channel dysfunction associated with BBMRs and schizophrenia	[118]	
	TASK5 (K _{2p} 15.1)	Therapeutic evaluation marker in male schizophrenia patients	[120]	
	TREK2 (K _{2p} 10.1)	Involvement in Schizophrenia development	[121]	
	TREK1 (K _{2p} 2.1)	Channel inhibition by SSRIs and genetic variants in MDD patients Channel inhibition in clinical anesthetic agents	[122]	
	TRESK (K _{2p} 18.1)	Channel dysfunction might increase the susceptibility to familial migraine	[123, 126]	
	TASK2 (K _{2p} 5.1)	Association with ion homeostasis and migraine	[128]	
	TALK1 (K _{2p} 16.1)	Genetic association with IGE	[129]	
Metabolic disorder	TALK1 (K _{2p} 16.1)	Channel genetic locus associated with T2D and MODY among different populations	[148, 149, 150]	
Kidney and urinary system disorders	TASK2 (K _{2p} 5.1)	Genetic mutation contributes to a renal disease, BEN	[16]	
	TREK1 (K _{2p} 2.1)	Reduced channel expression in DO and LUTS	[161, 164]	
GI disorders	TREK1 (K _{2p} 2.1)		[168]	
	TAAK (K _{2p} 4.1)	Downregulation in HSCR patient samples	[169]	

Table 3. Pathophysiological contributions of K_{2p} channels to human diseases The table lists currently-known mechanisms of K_{2p} channels that lead to different pathological conditions. The general pathomechanisms include dysfunction in cell proliferation, volume homeostasis, barrier integrity, endocrine signaling, and electrical activities in both neurons and smooth muscle cells. MS: multiple sclerosis, RA: rheumatoid arthritis, SCD: sickle cell disease, FHEIG: facial dysmorphism, hypertrichosis, epilepsy, intellectual/developmental delay, and gingival overgrowth syndrome, HSCR: Hirschsprung disease, T2D: type 2 diabetes, IBS: irritable bowel syndrome. CNS: central nervous system, GI: gastrointestinal system

Pathomechanism	Disease type	Disease name
Aberrant cell proliferation, differentiation and activation	Oncology	Cancer
	Autoimmune disease	MS and RA
	Neurodevelopmental disorder	FHEIG
Impaired volume regulation	Hematological disease	SCD
	Autoimmune disease	MS
Dysfunction in endothelial/ epithelial barrier integrity	GI disorder	HSCR
	CNS disease	Depression
Altered intracellular endocrine signaling	Metabolic disease	Schizophrenia
	CNS disorder	T2D
	CNS disorder	Pain/migraine epilepsy
Imbalanced electrochemical activities and dysregulated cellular excitability	Cardiovascular disorder	Arrhythmias
	GI disorder	IBS
	Kidney and urinary disorder	Detrusor overactivity

Hepatocellular carcinoma. With the help of bioinformatics and biochemical assays, downregulation of *KCNK5*, *KCNK17*, and *KCNK2* was observed in hepatocellular carcinoma (HCC) specimens, suggesting K_{2p} channels as a diagnostic biomarker for HCC [37].

Other cancers. Demethylation of long non-coding RNA (lncRNAs) in *KCNK15* and *WISP2* antisense *RNA 1 (KCNK15-AS1)* was observed in pancreatic cancer specimens, indicating a correlation between epigenetic changes in *KCNK15* and pancreatic carcinogenesis and metastasis [38]. Also, similar demethylation was found in gastric cancer patient samples and was due to inhibition of DNA methyltransferase 1-mitogen-activation protein kinase (DMNT1-MAPK) and histone deacetyltransferase 1-AKT (HDAC1-AKT) [39]. Moreover, the *KCNK15-AS1* lncRNA axis was found to contribute to the tumor progression in lung adenocarcinoma tissues via MicroRNA-202 (miR-202) and miR-307 [40]. These results indicate that CpG island methylation and histone acetylation might be common mechanisms of *KCNK15* modulation in post-transcription of oncogenes and tumor-suppressor genes [39] in several cancers despite different tumor microenvironments.

In addition to *KCNK15*, *KCNK5* upregulation was observed in specimens from a systemic screening of non-small-cell lung cancer (NSCLC) and pancreatic cancer [29]. TASK3 is recognized as an oxygen-sensing potassium channel in the human lung cancer cell line H146, suggesting TASK3 as a potential modulator of solid tumor formation [40, 41]. Also, strong TASK3 immunoactivity was observed in human gliomas specimens and TASK3 channel demonstrated functional relevance to isoflurane-induced cell death in U373 and LN393 human cell lines [42]. *KCNK2* upregulation was found in the human prostate cancer cell lines PC3 and LNCaP due to increased cell proliferation [43]. *KCNK6* was upregulated in cells from ovarian cancer patients, while it was downregulated in patients with colorectal and esophageal cancers. In CNS cancers, *KCNK12* downregulation was observed in astrocytoma and glioblastoma specimen [13, 28, 29].

Key mechanistic studies

The estrogen receptor α was found to mediate cell cycle checkpoints in cancer and cell proliferation by elevating *KCNK5* expression in the breast cancer cell lines MCF-7 and T47D [44]. Also, *KCNK9* upregulation promoted solid breast tumor formation due to its resistance to hypoxia and serum deprivation in C8 mouse embryonic fibroblast cells and Mus musculus mammary gland (NmuMG) epithelial cell-transferred mice [28]. As proof of concept, TASK3 blocking agents (zinc and methanandamide) induced cell apoptosis and reduced cell proliferation in the ovarian cancer cell lines SKOV-3 and OVCAR-3 [20]. A TASK3 monoclonal antibody was proven to reduce tumor growth due to channel internalization and dysfunctions. In addition to breast cancer, TASK3 genetic knockdown was found to dampen tumor invasion in the human gastric cancer cell lines KAYO-III and MKN-45. BL1249, a TREK1 activator, was identified to inhibit tumor proliferation and migration via hyperpolarization in the human pancreatic ductal adenocarcinoma cell line BxPC-3 [45]. Several studies have also demonstrated the importance of intracellular potassium levels in T cell effector functions, tumor clearance, and cell survival through protein phosphatase A (PP2A)-mediated Akt-mTOR phosphorylation [46], indicating K_{2p} channels as a promising candidate for anti-tumor effects in cancer therapy [31, 47].

K_{2p} channels in autoimmune diseases

Background

Autoimmunity is characterized by a loss of immune system self-tolerance towards own healthy cells and tissues. The etiology is only sparsely understood, however genetic (loss and gain of function mutations) and environmental factors (autoantibodies, UV exposure, and gut microbiome) are critically involved in disease development [48]. Altered expression and dysfunction of potassium channels in autoimmune diseases indicate a potential role in the disease pathology. For example, elevated Kv1.3 expression has been shown in autoreactive T cells from patients with type 1 diabetes, multiple sclerosis (MS), and rheumatoid arthritis [49, 50].

Also, the upregulation of Kv.7 was found in ulcerative HSCR patients, contributing to basolateral conductance [51]. In general, potassium channels contribute to the inflammatory responses in autoimmune diseases by regulating hyperpolarization-induced calcium influx together with Ca^{2+} release-activated Ca^{2+} (CRAC) channels and stromal interaction molecules (STIM) [52].

Human data

Reduced TASK1 and TASK3 channel expression were found in the inflammatory lesions of MS patients, particularly in CD11b⁺ macrophages and granulocytes. TASK channels might regulate cell apoptosis by initiating apoptotic volume decrease (AVD) and reducing the inhibition towards pro-apoptotic enzymes [19]. On the other hand, *KCNK9* upregulation was found in the colon of ulcerative colitis (UC) patients, implicating the contribution of *KCNK9* genetic variants in UC pathogenesis [53]. TASK2 upregulation in CD8⁺ T lymphocytes was observed in the CNS of relapsing-remitting multiple sclerosis (RRMS) patients in both acute and chronic phases. Correspondingly, TASK2 blockers and siRNA both confirmed their therapeutic potential in MS by modulating T cell effector functions [18]. In addition to findings in MS, comparable TASK2 upregulation was discovered in the blood- and synovial fluid-derived CD4⁺ T cells from RA patients and positively correlates with the DAS28 scores [54]. These findings indicate that TASK channel modulation might be beneficial to autoimmune diseases due to potassium-mediated T cell inflammatory responses.

Key mechanistic study

In the EAE model, TASK1 and TASK3 channels were found to mediate T cell proliferation and cytokine production by regulating intracellular calcium concentration via hyperpolarization [19]. Also, TASK1 was found to regulate oligodendrocyte differentiation *in vitro* and myelination *in vivo* via LINGO-1/ WNK1 phosphorylation [55]. TASK3 was discovered to modulate cell apoptosis and neurodegeneration via inflammation-mediated TNF- α activation [56]. Beyond MS pathology, TASK2-mediated intracellular Ca²⁺ signaling alternations in T cell subsets was observed in a mouse model of inflammatory bowel disease (IBD) [57]. KCNK2 downregulation on mRNA level was identified in murine dorsal root ganglia (DRG) neurons, leading to increased colon mechanosensitivity and UC development [58]. Also, TASK1 inhibition (anandamide and A293) was beneficial to EAE due to reduced calcium-dependent T cell activation, proliferation, and cytokine production [17, 19, 59]. However, TASK2 deficiency in mice showed no impact on EAE pathology due to the compensatory effects of TASK1 and Kv1.3 [60]. On the contrary, TREK1^{-/-} mice demonstrated exacerbated EAE phenotypes accompanied by elevated CNS T cell infiltration, altered endothelium integrity, and higher expression of adhesion molecules (VCAM1, ICAM1, and PECAM1) [61]. Similarly, suppressed TREK1 expression in intestinal epithelial cells worsened the colon inflammation by disrupting barrier integrity via histone deacetylation 1 (HDAC) and p38/mitogen-activated protein kinase (MAPK) pathway [62]. In summary, K_{2p} channels are involved in the development of autoimmune diseases by modulating calcium-mediated T cell activation, loss of barrier integrity, cell apoptosis, and eventually neurodegeneration.

K_{2p} channels in hematologic diseases

Background

Hematologic diseases affect blood and blood-producing organs and comprise anemias, sickle cell disease, glucose-6-phosphate dehydrogenase deficiency, and coagulopathies [63, 64]. Sickle cell disease (SCD) is one of the most common inherited hematologic diseases in regions affected by malaria worldwide and is caused by a genetic mutation in β -globin. SCD patients are characterized by sickle hemoglobin (HbS) and compromised red blood cell functions. Calcium-activated potassium channels (Gardos channels, KCNN4) contribute to the loss of potassium and erythrocyte dehydration in SCD. Inhibition of Gardos channels proved to alleviate red blood cell (RBC) dehydration and showed anti-sickling effects [65]. Also, a genetic mutation in Gardos channels contributes to rare anemias. For instance, R352H in KCNN4 was associated with decreased K⁺ content, increased conductance, and cell dehydration in hereditary xerocytosis [66, 67]. On the other hand, dehydration also occurred due to the loss of K⁺ via KCl channels [68], indicating a central role of potassium channels in the development of hematological diseases.

Human data

KCNK6, as a member of the K_{2p} channel family, was highly expressed in CD34⁺ cells and identified as a disease-contributing gene in the inherited polymorphisms (SNPs) of SCD patients from a Genome-wide Association Study (GWAS). *KCNK6* might regulate the disease pathogenesis by affecting erythroid differentiation and vaso-occlusive processes in SCD [69].

Key mechanistic study

Early in the 1990s, scientists suggested the association of cation ions with SCD [70, 71], particularly potassium loss in RBC dehydration [72, 73]. TWIK2 channels are highly expressed in CD34⁺ stem cells, the cell population associated with erythroid differentiation. Besides, the potassium homeostasis of erythrocytes is critical for HbS polymerization, RBC hemolysis, and the development of SCD [69]. However, the detailed mechanism of TWIK2 channel contribution to SCD disease is still unclear and requires further mechanistic studies.

K_{2p} channels in cardiovascular disorders

Background

Cardiovascular diseases include blood vessel blockage, myocardial dysfunctions, and arrhythmias [74, 75]. For example, ischemic stroke is associated with blood clot formation and vessel blockage [76]. Diseases with myocardial dysfunctions include atherosclerosis, coronary arterial disease (CAD), myocardial infarction (MI), and left ventricular (LV) dysfunctions [77, 78]. Arrhythmias refer to disorganized electrical activities in the cardiac system such as tachycardia, idiopathic ventricular fibrillation (IVF), atrial fibrillation (AF), and congenital long QT syndrome (LQTS) [79, 80].

The physiological functions of the cardiovascular system rely on depolarization-mediated smooth muscle tone for which K_{2p} channels play a critical role by coordinating voltage-gated calcium channels (VGCC) and stabilizing the membrane potential [82]. Hence, K_{2p} channels contribute to the development of cardiovascular diseases.

Human data

In the 2000s, a growing number of K_{2p} channel studies related to the cardiovascular system were conducted based on the high expression in pulmonary artery smooth muscle cells (PASMC) [82, 83]. In addition to the high expression, several K_{2p} channels are associated with the development of cardiovascular disorders. For instance, *KCNK17* genetic variants were associated with vessel blockage, susceptibility to ischemic stroke in Caucasian, and cerebral hemorrhage in Chinese populations [38, 84, 85]. In addition to *KCNK17*, multiple *KCNK5* genetic variants were found to be a common risk factor of CAD and migraine, suggesting overlapping mechanisms in disease pathogenesis [86, 87]. Also, the *TASK2* genetic variant rs10947789 was discovered as an overlapping risk gene in the blood samples of CAD and MI patients and was associated with increased platelet counts and volume [88]. Apart from being a risk factor, *TASK2* channels regulated myocardial functions by modulating membrane potentials and respiration via oxygen chemosensitivity [89].

TASK1 channels are a common molecular target for treating arrhythmias (amiodarone, vernakalant, flecainide, and carvedilol), and A293, a potent and specific *TASK1* blocker, displayed potent antiarrhythmic effects [90, 91]. Also, *TASK1* inhibition was found to attenuate cardiovascular dysfunctions in AF patients by prolonging action potentials [92, 93], and *TASK1* missense mutation and reduced currents were observed in patients with pulmonary arterial hypertension [94, 95]. As a proof of concept, phospholipase inhibitor ONO-RS-082 was able to increase *TASK1* currents in human PASMC via channel activation [96]. Also, decreased atrial *KCNK3* expression was observed in patients with left ventricular (LV) dysfunction but was increased in chronic AF (cAF) patients. *KCNK3* channel inhibition prolonged action potential (AP) duration and showed beneficial effects in cAF patients [97]. Therefore, *TASK1* inhibitors and activators both might be beneficial for treating cardiovascular disorders depending on the different pathological conditions. In whole-exome sequencing (WES) research, mechanosensitive *TREK1* and *TREK2* channels are highly expressed throughout the cardiac system. Atrial and ventricular *TREK1* downregulation was found in AF patients and contributed to cardiac rhythmic regulation [97]. Also, a later study confirmed significant mRNA reduction (-80%) in the atrium of AF and HF patients, leading to prolonged atrial effective refractory periods [97, 98]. In whole blood samples, a point mutation in *TREK1* increased potassium permeability, mechano-sensitivity, and contributed to right ventricular outflow tract (RVOT) tachycardia [99]. G88R mutation and genetic variants in *KCNK17* were discovered in whole blood samples and induced pluripotent stem cell-derived cardiomyocytes to enhance channel currents, hyperpolarization, and contribute to the pathologies of IVF and LQTS [100, 101]. Moreover, *KCNK17* was found to be sensitive to antiarrhythmic drugs and involved in drug working mechanisms [102]. On the other hand, reduced *TALK2* currents in atria and ventricles were observed in patients with heart failure (HF) and atrial fibrillation [103]. Therefore, *TALK2* activators and blockers both showed therapeutic potentials in treating arrhythmias and heart failure. In summary, K_{2p} channels are a promising target for treating a range of cardiovascular disorders.

Key mechanistic study

Although the mechanism of how *KCNK17* modulate the susceptibility to ischemic stroke is still unclear, TREK1 silencing by short-hairpin RNA displayed protective effects in rat cardiomyocytes by decreasing cell apoptosis under ischemic injury conditions [104]. Also, in the transient middle cerebral artery occlusion (tMCAO) model, *TASK1*^{-/-} mice displayed larger infarcted volumes, but *TRAAK*^{-/-} mice showed preserved brain metabolism and pH. Therefore, *TASK1* channel-induced hyperpolarization might be an intrinsic defense mechanism, while high levels of organic osmolytes in *TRAAK*^{-/-} mice avoided ischemic-induced cell death [105, 106].

For K_{2p} channel modulation in myocardial functions, a naïve TREK-1-like current discovered in rat ventricular cardiomyocytes is activated by ATP through cytosolic phospholipase A_2 , cAMP-mediated protein kinase A (PKA), and tyrosine kinase pathways [107]. Also, endothelin-1 (ET-1) was discovered as a TREK1 upstream regulator in calcium-mediated vasoconstriction via Gq protein-coupled protein kinase C (PKC) signaling [108]. β (IV) spectrin, an actin-associated protein, was found to regulate TREK1 membrane trafficking by colocalization with the channel [109, 110]. Therefore, TREK1 channels might coordinate with G protein coupled receptors and its downstream effector proteins to regulate myocardial functions under physiological conditions. On the other hand, histone deacetylase (HDAC) inhibitors increased TREK1 expression and prolonged action potential duration in murine atrial cardiomyocytes, indicating that epigenetic changes also modulate TREK1 functions [111].

Although atrial *KCNK2* expression was reduced in the AF mouse model, TREK1 specific deletion in fibroblasts attenuated cardiac fibrosis and dysfunctions through PKC-mediated oxidative stress and Jun N-terminal kinase (JNK)-mediated cell death [97, 112]. Besides, reduced atrial *TASK1* expression was observed in an AF mouse model (*CREM* transgenic mice) and a HF disease model (transverse aortic constriction) [113]. Furthermore, in *TASK1*^{-/-} mice, prolonged action potential and QT intervals were observed in electrocardiograms, accompanied by reduced autonomic variability and sympathetic overactivity [114, 115]. However, there is currently no genetic knockout mice study or mechanistic data for the role of *TASK2* channels in cardiovascular disorders. Propafenone, a commonly used antiarrhythmic drug, was found to activate the *TALK2* channel with a 7.8-fold current increase in mammalian Chinese hamster ovary (CHO) cells. Also, *TALK2* expressed in *Xenopus* oocytes showed sensitivity to most antiarrhythmic drugs, such as propafenone, quinidine, mexiletine, and metoprolol [102]. However, mechanistic studies of *TALK2* in arrhythmias are largely missing.

K_{2p} channels in CNS diseases

Background

Central nervous system (CNS) diseases include disorders of the brain, spinal cord, and nerves. Neurodevelopmental diseases affect CNS development, where mutations lead to abnormal brain size and dysfunctions. Psychiatric disorders are brain disorders characterized by abnormal mental and behavioral phenotypes. Further CNS-related diseases such as pain, migraine, and epilepsy are based on neuronal hyperexcitability, abnormal biochemical metabolisms, and aggregations. Potassium channels are a critical modulator of electrochemical and ion homeostasis, and the high expression and central functions of K_{2p} channels in the CNS argue for an essential role in CNS diseases.

Human data

Neurodevelopmental disorders are mainly caused by familial mutations. For instance, FHEIG (Facial dysmorphism, Hypertrichosis, Epilepsy, Intellectual disability, Gingival overgrowth) is related to the *KCNK4* missense mutation negatively affecting lateral intramembrane fenestration during CNS development [116]. Birk Barel Mental Retardation Syndrome (BBMRS) is a maternally transferred disease characterized by mental retardation,

hypotonia, and dimorphisms and caused by a *KCNK9* missense mutation leading to impaired channel functions. Flufenamic acid (FFA) application was found to enhance TASK3 channel currents and improve the symptoms in younger BBMRs patients [117].

Emerging evidence also suggests a role of TASK3 channels in psychiatric disorders. For example, rs4736253a, a genetic locus near *KCNK9*, was associated with schizophrenia [118]. Furthermore, hypomethylated *KCNK15* was associated with remission in male schizophrenia patients [119]. Similarly, demethylation in *KCNK10* was associated with neuronal growth and cerebellum development in schizophrenia patients [120]. Genetic variants in *KCNK2* were identified to influence treatment resistance in patients with major depressive disorders (MDD) [121].

Migraine, pain, and epilepsy are related to neuronal hyperexcitability. In migraine with and without aura (MA and MO), several *KCNK18* mutations were associated with reduced neuronal current threshold and high spike frequencies [122]. Some contradictory studies discovered that the *KCNK18* variants A34R and C110R were detectable in both migraine patients and controls [123, 124], suggesting that a single TRESK mutation is not sufficient to cause migraine. Afterward, the C110R variant in TRESK was confirmed to show preserved currents in human nociceptors, and only the frameshift mutation F139WfsX24 led to a loss of TRESK function [125]. Bupivacaine, a clinical anesthetic agent, was found to inhibit TREK1 channels. Also, Aristolochic acid (AristA), a plant extract medicine to treat pain, enhanced TREK1 and TREK2 currents but inhibited TRESK currents [126]. *KCNK5* was also identified as a risk factor for migraine from a GWAS [127], whereas *KCNK16* and *KCNK17* variants were suggested to be risk factors for idiopathic generalized epilepsy (IGE) due to altered channel currents and spike frequencies [128].

Key mechanistic study

There is currently no mechanistic study of K_{2p} channels in neurodevelopmental disorders. However, potential mechanisms of K_{2p} channels contributing to psychiatric disorders were suggested. For example, a TREK2 mutation in the protein kinase A (PKA) phosphorylation site abolished the norepinephrine-mediated suppression of neuronal excitability and the development of schizophrenia [129]. Besides, selective serotonin reuptake inhibitors (SSRIs), a group of commonly used antidepressants, are effective through TREK1 channel inhibition, among other mechanisms [130, 131]. Correspondingly, a depression-resistant phenotype was observed in TREK1^{-/-} mice due to higher efficiency of 5-HT neurotransmission and reduced stress-mediated corticosterone levels in serum [132, 133]. In addition, spadin, a specific TREK1 blocker, was discovered to show anti-depressive effects by activating the 5-HT1A receptor, the cAMP-response element (CREB), brain-derived neurotrophic factor (BDNF) signaling, and hippocampal neurogenesis [134, 135]. In contrast, ostruthin, an element extracted from plants, showed anxiolytic and anti-depressive effects by activating TREK1 channels, increasing channel currents, and reducing stress-mediated c-Fos signaling [136].

Also, TREK1^{-/-} mice displayed hyperalgesia toward mechanical and thermal stimuli [137], while μ -opioid receptor-mediated TREK1 activation showed morphine-mediated analgesic effects without opioid-induced adverse effects [138]. Further, 11-Deoxy prostaglandin F2 α , a TREK2 selective activator, showed analgesic effects by reducing the calcium influx in mouse primary dorsal root ganglia (DRG) [139], whereas TREK2 downregulation by siRNA induced depolarization of the nociceptors in DRG neurons and exacerbated hyperalgesia in rats [140]. Similarly, TRESK mutations in mouse trigeminal ganglion (TG) cells showed lower current threshold among action potential initiation, increased spike frequencies, and increased migraine susceptibility [141], whereas TRESK overexpression led to reduced spike formation and excitability [142].

K_{2p} channels in metabolic disorders

Background

Metabolic disorder refers to a deficiency in enzymes during metabolic processes, e.g., the metabolism of glucose, carbohydrates, amino acids, and fatty acids. Type 2 diabetes (T2D) is a common metabolic disorder associated with family history, chronic diseases, age, and obesity and is characterized by disrupted glucose-mediated insulin secretion. The correlation between potassium and glucose metabolism has been proposed since the late 1900s. For instance, potassium depletion leads to impairment in insulin secretion and glucose tolerance through depolarization [143, 144], while higher potassium intake reduces the risk of T2D development [145, 146].

Human data

In pancreatic beta cells, *KCNK16* was identified as susceptibility locus of T2D from several GWAS in East Asian, Indian, and European populations and is involved in pancreatic β cell development and insulin secretion [147, 148]. Moreover, the association of *KCNK16* with T2D was strengthened from the results of small RNA sequencing [149]. Lately, a *KCNK16* gain of function was also identified in patients with maturity-onset diabetes of the young (MODY) by affecting calcium signaling and glucose-stimulated insulin secretion (GSIS) [150].

Key mechanistic study

Enriched TASK1 channel expression was found in the plasma membrane of β cells in both humans and rodents. TASK1 can regulate hyperpolarization by interacting with voltage-dependent calcium channels (VDCC), suggesting TASK1 as a modulator of calcium signaling and the development of T2D. As a proof of concept, TASK1 conditional deletion in pancreatic β cells led to increased glucose-stimulated depolarization, GSIS, and improved glucose tolerance [151].

The genetic polymorphism of TALK1 channel rs1535500 was associated with T2D. In *TALK1*^{-/-} mice, increased β -cell depolarization, enhanced GSIS reduced calcium-mediated ER stress, and islet dysfunction were observed [152, 153]. Besides, cytokine-mediated TALK1 inhibition showed protective effects on β -cells by facilitating calcium influx and GSIS under inflammation [1554, indicating modulatory functions of K_{2p} channels in glucose tolerance and T2D development.

K_{2p} channels in kidney and urinary system disorders

Background

Kidney and urinary-tract disorders include kidney dysfunctions, abnormal urinary filtration, and urination. Potassium channels are central modulators of resting membrane potential in smooth muscle cells, renal vascular cell contractility, and ion homeostasis, supporting a critical role in various human kidney and urinary system disorders [155, 156]. For instance, reduced urinary potassium excretion is associated with reduced renal mass and dysfunction in glomerular filtration [157]. On the contrary, elevated urinary potassium excretion and high potassium diet lowered the risk of chronic kidney disease development [158].

Human data

pH-sensitive TASK2 channels are highly abundant in the nephron of human kidneys, especially in tubular epithelia, and are inhibited by external acidic pH [159]. Correspondingly, T108P, a missense variant in *KCNK5* leading to a loss of channel function, is associated with Balkan endemic nephropathy (BEN), a familial and chronic kidney disease [16].

For TREK1 channels, downregulated expression and reduced currents were observed in human detrusor overactivity (DO), an abnormal response of the bladder to physiological

stretches. Furthermore, DO myocytes failed to dilate while exposed to TREK1 channel openers, which supports the role of TREK1 in regulating bladder contraction by interacting with cytoskeletal proteins [160]. In addition, diminished TREK1 expression was also found in the SNPs of patients with lower urinary tract symptoms (LUTS) and was associated with urinary defects [161].

Key mechanistic study

In TASK2 deficient mice, the pH and concentration of HCO_3^- was reduced in the blood but increased in the urinary system, indicating channel inhibition and metabolic acidosis due to renal bicarbonate loss [162]. In TREK1^{-/-} mice, elevated muscle tone and more contractile force in response to stimulations were found in myocytes. Also, TREK1^{-/-} animals revealed increased micturition durations and bladder capacity. However, a mixed effect was observed in global knockout mice, suggesting that further studies are required with conditional knockout animals [163]. Besides, significant TREK1 downregulation was found in the obstructor myocytes of the DO mouse model after bladder obstruction [164], whereas channel upregulation was observed in the rat model [165], indicating the urgent need of a promising DO animal model for representing human patients.

K_{2p} channels in gastrointestinal (GI) disorders

Background

Gastrointestinal (GI) disorder refers to defects in GI tracts, including the esophagus, stomach, intestines, and rectum. Potassium homeostasis is one vital factor, maintaining the physiological functions of muscle tone and motility of GI tracts. Particularly the mechanosensitive K_{2p} channels are highly expressed in mechanosensory smooth muscle cells and Cajal cells in the human GI system [58, 166, 167], suggesting a potential for K_{2p} channel modulation in the GI system.

Human data

Hirschsprung disease (HSCR) is a congenital disorder within the GI system due to reduced TREK1 expression and missing intestinal aganglionic and ganglionic neurons [168]. Also, TRAAK downregulation was observed in aganglionic and ganglionic neurons in the colon, indicating disrupted K_{2p} channel functions for maintaining epithelium barrier integrity and contributing to the disease development of HSCR [169]. The role of mechanosensitive K_{2p} channels in other GI disorders remains unknown and requires further investigations.

Key mechanistic study

The association of mechanosensitive K_{2p} channels with GI disorders was discovered in the context of high channel expression in murine small and large intestines [170]. For example, abundant TASK2 expression was found in the murine intestinal epithelium and played a critical role in maintaining the anion and fluid secretions. Tetrapentylammonium, a TASK2 inhibitor, was found to abolish the anion secretory current [171]. Besides, altered potassium channel-mediated muscle contraction induces irritable bowel syndrome (IBS) [172]. Riluzole, a mechanosensitive channel activator, reduced the hypercontractility of colon myocytes, indicating TREK1 as a potential therapeutic target for the hypercontraction in IBS [167]. Although it is known now that mechano-gated K_{2p} channels regulate the GI system by maintaining epithelial barrier integrity and smooth muscle tone contraction, the underlying molecular mechanisms and related signaling pathways are still unclear.

Outlook

The growing research on K_{2p} channels not only brings new insights into the field but also reveals obstacles for both researchers and physicians. The major difficulties include translational roadblocks, lack of modulators with high specificity, and risk of adverse effects. There are already several examples of failed translation in K_{2p} channel research: TASK1 deficient mice showed reduced T cell effector functions and ameliorated motor dysfunction, whereas TASK1 downregulation was found in MS patients with elevated inflammatory responses. Moreover, TASK2 upregulation was observed in both RA and MS patients; however, TASK2^{-/-} mice showed a EAE disease course comparable with WT. The usage of patient-derived and -induced pluripotent stem cells and computer-based prediction models might be helpful to overcome the translational roadblock. Nonspecific channel modulators and serious adverse effects are further problems hampering human application, especially regarding drug development for K_{2p} channels. For instance, the commonly used anesthetic bupivacaine blocks not only TASK1 and TASK3 but also TREK1 [173, 174]. TASK2 inhibition via class III antiarrhythmic drugs is beneficial for cardiovascular disorders but associated with potential adverse effects such as renal failure [175]. Structural studies of K_{2p} channels on the allosteric ligand-binding site might help to identify more specific pharmacological modulators. Also, detailed studies on K_{2p} channel expression and function in different tissues might help to predict adverse events. Besides overcoming current obstacles for human application with the well-studied plasma membrane-localized K_{2p} channel, further research on intracellular K_{2p} channels (e.g., THIK2 in the endoplasmic reticulum, TWIK1 in endosomes, and TWIK2 in lysosomes) [176, 177] might open up new therapeutic avenues due to novel targets in the channel trafficking process. For example, TWIK2^{-/-} mice demonstrated pulmonary hypertension with altered vasocontractility [178, 179], and TWIK2 deficient macrophages prevented pulmonary inflammation in mice [180]. Overall, the K_{2p} research field is indispensable and promises a deeper understanding of the pathophysiology of human disorders, allowing us to develop new diagnostic and therapeutic strategies.

Conclusion

Solid evidence suggests that K_{2p} channels are major diagnostic and therapeutic candidates for several human diseases. However, the insights into the underlying molecular processes are only fragmentary. Nevertheless, common pathophysiological mechanisms can be identified as follows (Table 3): (1) aberrant cell proliferation, differentiation, and activation are associated with cancer, neurodevelopmental disease (FHEIG), and autoimmunity; (2) impaired volume regulation is associated with erythrocyte abnormalities in sickle cell disease; (3) dysfunction of endothelial/epithelial barrier integrity in brain and colon is associated with MS and HSCR; (4) altered intracellular and endocrine signaling are found in depression, schizophrenia, and type 2 diabetes; and (5) imbalanced electrochemical activities and dysregulated cellular excitability are associated with neuropathic pain, migraine, epilepsy, and cardiac arrhythmias as well as smooth muscle dysfunction, e.g., in irritable bowel disease and detrusor hyperactivity. However, to warrant further translational research and to develop applications for human diseases, deeper insight into the underlying molecular processes are required.

Acknowledgements

The authors give their special thanks to the people listed below for proofreading the manuscript: Dr. I-Na Lu, Dr. Anna Speicher, Jolien Wolbert and Laura Vinnenberg.

Author Contributions

S.M and T.R contributed to the conception and structure of the manuscript discussing the involvement of K_{2p} channels in human diseases. LM. L collected the relevant papers, covering the topic in the field, interpreted, and providing answers and suggestions to the future K_{2p} channel study in human diseases. LM. L drafted the manuscript with supports of critical revising from T.M in oncology section and T.B. for K_{2p} channel physiology. Finally, T.R and S.M provided critical revision of the article and approved the final version to be submitted.

Funding Sources

This work was funded by the “Else Kröner-Fresenius-Stiftung” (2018_A03 to T.R.) and by the “Innovative Medizinische Forschung (IMF)” Münster (I-RU211811 to T.R.). The authors received further funding resources from CiM-IMPRS Graduate Program and Interdisziplinäres Zentrum für Klinische Forschung (IZKF) in Münster with project number: Meu3/015/18 and ONO Pharmaceuticals Co. Ltd. specifically in the project of the role of TWIK2 channel in a mouse model of neuroinflammation.

Disclosure Statement

The authors have no conflicts of interest to declare in this review article.

References

- 1 Lesage F, Guillemare E, Fink M, Duprat F, Lazdunski M, Romey G, et al.: TWIK-1, a ubiquitous human weakly inward rectifying K⁺ channel with a novel structure. *EMBO J* 1996;15:1004–1011.
- 2 Gada K, Plant LD: Two-pore domain potassium channels: emerging targets for novel analgesic drugs: *IUPHAR Review* 26. *Br J Pharmacol* 2019;176:256–266.
- 3 Hughes S, Foster RG, Peirson SN, Hankins MW: Expression and localisation of two-pore domain (K2P) background leak potassium ion channels in the mouse retina. *Sci Rep* 2017;7:1–14.
- 4 Schewe M, Nematian-Ardestani E, Sun H, Musinszki M, Cordeiro S, Bucci G, et al.: A Non-canonical Voltage-Sensing Mechanism Controls Gating in K2P K⁺ Channels. *Cell* 2016;164:937–949.
- 5 Lotshaw DP: Biophysical, pharmacological, and functional characteristics of cloned and native mammalian two-pore domain K⁺ channels. *Cell Biochem Biophys* 2007;47:209–256.
- 6 O’Connell AD, Morton MJ, Hunter M: Two-pore domain K⁺ channels - Molecular sensors. *Biochim Biophys Acta - Biomembr* 2002;1566:152–161.
- 7 Enyedi P, Czirják G: Molecular background of leak K⁺ currents: Two-pore domain potassium channels. *Physiol Rev* 2010;90:559–605.
- 8 Franks NP, Honoré E: The TREK K_{2p} channels and their role in general anaesthesia and neuroprotection. *Trends Pharmacol Sci* 2004;25:601–608.
- 9 Schmidt C, Wiedmann F, Kallenberger SM, Ratte A, Schulte JS, Scholz B, et al.: Stretch-activated two-pore-domain (K2P) potassium channels in the heart: Focus on atrial fibrillation and heart failure. *Prog Biophys Mol Biol* 2017;130:233–243.
- 10 Steinberg EA, Wafford KA, Brickley SG, Franks NP, Wisden W: The role of K2P channels in anaesthesia and sleep. *Pflügers Arch Eur J Physiol* 2015;467:907–916.
- 11 Li XY, Toyoda H: Role of leak potassium channels in pain signaling. *Brain Res Bull* 2015;119:73–79.
- 12 Hancox JC, James AF, Marrion NV, Zhang H, Thomas D: Novel ion channel targets in atrial fibrillation. *Expert Opin Ther Targets* 2016;20:947–958.
- 13 Williams S, Bateman A, O’Kelly I: Altered Expression of Two-Pore Domain Potassium (K2P) Channels in Cancer. *PLoS One* 2013;8:e74859.
- 14 Bittner S, Ruck T, Schuhmann MK, Herrmann AM, Maati HMO, Bobak N, et al.: Endothelial TWIK-related potassium channel-1 (TREK1) regulates immune-cell trafficking into the CNS. *Nat Med* 2013;19:1161–1165.

- 15 Ehling P, Cerina M, Budde T, Meuth SG, Bittner S: The CNS under pathophysiologic attack—examining the role of K2P channels. *Pflügers Arch Eur J Physiol* 2015;467:959–972.
- 16 Reed AP, Bucci G, Abd-Wahab F, Tucker SJ: Dominant-negative effect of a missense variant in the TASK-2 (KCNK5) K⁺ channel associated with Balkan Endemic Nephropathy. *PLoS One* 2016;11:1–12.
- 17 Bittner S, Meuth SG, Göbel K, Melzer N, Herrmann AM, Simon OJ, et al.: TASK1 modulates inflammation and neurodegeneration in autoimmune inflammation of the central nervous system. *Brain* 2009;132:2501–2516.
- 18 Bittner S, Bobak N, Herrmann AM, Göbel K, Meuth P, Höhn KG, et al.: Upregulation of K2P5.1 potassium channels in multiple sclerosis. *Ann Neurol* 2010;68:58–69.
- 19 Meuth SG, Bittner S, Meuth P, Simon OJ, Budde T, Wiendl H: TWIK-related acid-sensitive K⁺ channel 1 (TASK1) and TASK3 critically influence T lymphocyte effector functions. *J Biol Chem* 2008a;283:14559–14570.
- 20 Anni I, Leigh J, Viren A, Gerhard VS, Aveil W, Daniel H, Anish B: Expression and Prognostic Significance of the Oncogenic K2P Potassium Channel KCNK9 (TASK-3) in Ovarian Carcinoma. *Anticancer Res* 2013;33:1401–1408.
- 21 Zúñiga R, Valenzuela C, Concha G, Brown N, Zúñiga L: TASK-3 downregulation triggers cellular senescence and growth inhibition in breast cancer cell lines. *Int J Mol Sci* 2018;19:1033.
- 22 William Audeh M: Genetic and environmental factors in cancer pathogenesis. *Princ Pract Surg Oncol A Multidiscip Approach to Difficult Probl* 2012;512:135–153.
- 23 Rudolph M, Anzeneder T, Schulz A, Beckmann G, Byrne AT, Jeffers M, et al.: AKT1E17K mutation profiling in breast cancer: Prevalence, concurrent oncogenic alterations, and blood-based detection. *BMC Cancer* 2016;16:1–12.
- 24 Sahar S, Sassone-Corsi P: Metabolism and cancer: The circadian clock connection. *Nat Rev Cancer* 2009;9:886–896.
- 25 Sherburn: Hull Royal Infirmary. a Case of Dislocation of the Wrist Backwards. *Lancet* 1889;133:985–986.
- 26 Thomas SM, Brugge JS: Cellular Functions Regulated By Src Family Kinases. *Annu Rev Cell Dev Biol* 1997;13:513–609.
- 27 Plummer HK, Dhar MS, Cekanova M, Schuller HM: Expression of G-protein inwardly rectifying potassium channels (GIRKs) in lung cancer cell lines. *BMC Cancer* 2005;5:1–10.
- 28 Dookeran KA, Auer P: The Emerging Role of Two-Pore Domain Potassium Channels in Breast Cancer. *J Glob Epidemiol Environ Health* 2017;2017:27–36.
- 29 Santarius T, Bignell GR, Greenman CD, Widaa S, Chen L, Mahoney CL, et al.: GLO1 — A Novel Amplified Gene in Human Cancer. *Genes Chromosomes Cancer* 2010;49:711–725.
- 30 Hammadi M, Chopin V, Matifat F, Dhennin-Duthille I, Chasseraud M, Sevestre H, et al.: Human ether à-gogo K⁺ channel 1 (hEag1) regulates MDA-MB-231 breast cancer cell migration through Orai1-dependent calcium entry. *J Cell Physiol* 2012;227:3837–3846.
- 31 Sun H, Luo L, Lal B, Ma X, Chen L, Hann CL, et al.: A monoclonal antibody against KCNK9 K⁺ channel extracellular domain inhibits tumour growth and metastasis. *Nat Commun* 2016;7:10339.
- 32 Wang L, Song L, Li J, Wang Y, Yang C, Kou X, et al.: Bone sialoprotein- α v β 3 integrin axis promotes breast cancer metastasis to the bone. *Cancer Sci* 2019;110:3157–3172.
- 33 Alan Harris R, Nagy-Szakal D, Kellermayer R: Human metastable epiallele candidates link to common disorders. *Epigenetics* 2013;8:157–163.
- 34 Sánchez-Miguel DS, García-Dolores F, Rosa Flores-Márquez M, Delgado-Enciso I, Pottosin I, Dobrovinskaya O: TRESK potassium channel in human T lymphoblasts. *Biochem Biophys Res Commun* 2013;434:273–279.
- 35 Pocsai K, Kosztka L, Bakondi G, Gönczi M, Fodor J, Dienes B, et al.: Melanoma cells exhibit strong intracellular TASK-3-specific immunopositivity in both tissue sections and cell culture. *Cell Mol Life Sci* 2006;63:2364–2376.
- 36 D'Arcangelo D, Scatozza F, Giampietri C, Marchetti P, Facchiano F, Facchiano A: Ion channel expression in human melanoma samples: In silico identification and experimental validation of molecular targets. *Cancers (Basel)* 2019;11:446.
- 37 Li WC, Xiong ZY, Huang PZ, Liao YJ, Li QX, Yao ZC, et al.: KCNK levels are prognostic and diagnostic markers for hepatocellular carcinoma. *Aging (Albany NY)* 2019;11:8169–8182.
- 38 He L, Ma Q, Wang Y, Liu X, Yuan Y, Zhang Y, et al.: Association of variants in KCNK17 gene with ischemic stroke and cerebral hemorrhage in a chinese population. *J Stroke Cerebrovasc Dis* 2014;23:2322–2327.

- 39 Zhang H, Zhang Z, Wang D: Epigenetic regulation of incRNA KCNK15-ASI in gastric cancer. *Cancer Manag Res* 2019;11:8589–8602.
- 40 Peng J, Chen XL, Cheng HZ, Xu ZY, Wang H, Shi ZZ, et al.: Silencing of KCNK15-AS1 inhibits lung cancer cell proliferation via upregulation of miR-202 and miR-370. *Oncol Lett* 2019;18:5968–5976.
- 41 Hartness ME, Lewis A, Searle GJ, O’Kelly I, Peers C, Kemp PJ: Combined Antisense and Pharmacological Approaches Implicate hTASK as an Airway O₂ Sensing K⁺ Channel. *J Biol Chem* 2001;276:26499–26508.
- 42 Meuth SG, Herrmann AM, Ip CW, Kanyshkova T, Bittner S, Weishaupt A, et al.: The two-pore domain potassium channel TASK3 functionally impacts glioma cell death. *J Neurooncol* 2008;87:263–270.
- 43 Voloshyna I, Besana A, Castillo M, Matos T, Weinstein IB, Mansukhani M, et al.: TREK-1 is a novel molecular target in prostate cancer. *Cancer Res* 2008;68:1197–1203.
- 44 Alvarez-Baron CP, Jonsson P, Thomas C, Dryer SE, Williams C: The two-pore domain potassium channel KCNK5: Induction by estrogen receptor α and role in proliferation of breast cancer cells. *Mol Endocrinol* 2011;25:1326–1336.
- 45 Sauter DRP, Sørensen CE, Rapedius M, Brüggemann A, Novak I: pH-sensitive K⁺ channel TREK-1 is a novel target in pancreatic cancer. *Biochim Biophys Acta* 2016;1862:1994–2003.
- 46 Eil R, Vodnala SK, Clever D, Klebanoff CA, Sukumar M, Pan JH, et al.: Ionic immune suppression within the tumour microenvironment limits T cell effector function. *Nature* 2016;537:539–543.
- 47 Cikutović-Molina R, Herrada AA, González W, Brown N, Zúñiga L: TASK-3 gene knockdown dampens invasion and migration and promotes apoptosis in KATO III and MKN-45 human gastric adenocarcinoma cell lines. *Int J Mol Sci* 2019;20:6077.
- 48 Theofilopoulos AN, Kono DH, Baccala R: The multiple pathways to autoimmunity. *Nat Immunol* 2017;18:716–724.
- 49 Wulff H, Calabresi PA, Allie R, Yun S, Pennington M, Beeton C, et al.: The voltage-gated Kv1.3 K⁺ channel in effector memory T cells as new target for MS. *J Clin Invest* 2003;111:1703–1713.
- 50 Beeton C, Wulff H, Standifer NE, Azam P, Mullen KM, Pennington MW, et al.: Kv1.3 channels are a therapeutic target for T cell-mediated autoimmune diseases. *Proc Natl Acad Sci U S A* 2006;103:17414–17419.
- 51 Al-Hazza A, Linley J, Aziz Q, Hunter M, Sandle G: Upregulation of basolateral small conductance potassium channels (KCNQ1/KCNE3) in ulcerative colitis. *Biochem Biophys Res Commun* 2016;470:473–478.
- 52 Feske S, Wulff H, Skolnik EY: Ion channels in innate and adaptive immunity. *Annu Rev Immunol* 2015;33:291–353.
- 53 Saadati HR, Wittig M, Helbig I, Häsler R, Anderson CA, Mathew CG, et al.: Genome-wide rare copy number variation screening in ulcerative colitis identifies potential susceptibility loci. *BMC Med Genet* 2016;17:1–10.
- 54 Bittner S, Bobak N, Feuchtenberger M, Herrmann AM, Göbel K, Kinne RW, et al.: Expression of K2P5.1 potassium channels on CD4⁺T lymphocytes correlates with disease activity in rheumatoid arthritis patients. *Arthritis Res Ther* 2011;13:R21.
- 55 Albrecht S, Korr S, Nowack L, Narayanan V, Starost L, Stortz F, et al.: The K2P -channel TASK1 affects Oligodendroglial differentiation but not myelin restoration. *Glia* 2019;67:870–883.
- 56 ElHachmane MF, Rees KA, Veale EL, Sumbayev VV, Mathie A: Enhancement of TWIK-related acid-sensitive potassium channel 3 (TASK3) two-pore domain potassium channel activity by tumor necrosis factor α . *J Biol Chem* 2014;289:1388–1401.
- 57 Nakakura S, Matsui M, Sato A, Ishii M, Endo K, Muragishi S, et al.: Pathophysiological significance of the two-pore domain K⁺ channel K2P5.1 in splenic CD4⁺CD25⁻ T cell subset from a chemically-induced murine inflammatory bowel disease model. *Front Physiol* 2015;6:1–10.
- 58 La JH, Gebhart GF: Colitis decreases mechanosensitive K_{2p} channel expression and function in mouse colon sensory neurons. *Am J Physiol - Gastrointest Liver Physiol* 2011;301:165–174.
- 59 Bittner S, Bauer MA, Ehling P, Bobak N, Breuer J, Herrmann AM, et al.: The TASK1 channel inhibitor A293 shows efficacy in a mouse model of multiple sclerosis. *Exp Neurol* 2012;238:149–155.
- 60 Bittner S, Bobak N, Hofmann MS, Schuhmann MK, Ruck T, Göbel K, et al.: Murine K2P5.1 deficiency has no impact on autoimmune neuroinflammation due to compensatory K2P3.1- and Kv1.3-dependent mechanisms. *Int J Mol Sci* 2015;16:16880–16896.
- 61 Bittner S, Ruck T, Fernández-Orth J, Meuth SG: TREK-king the blood-brain-barrier. *J Neuroimmune Pharmacol* 2014;9:293–301.

- 62 Huang H, Liu JQ, Yu Y, Mo LH, Ge RT, Zhang HP, et al.: Regulation of TWIK-related potassium channel-1 (Trek1) restitutes intestinal epithelial barrier function. *Cell Mol Immunol* 2016;13:110-118.
- 63 Lang K, Roll B, Myssina S, Schittenhelm M, Scheel-Walter HG, Kanz L, et al.: Enhanced erythrocyte apoptosis in sickle cell anemia, thalassemia and glucose-6-phosphate dehydrogenase deficiency. *Cell Physiol Biochem* 2002;12:365-372.
- 64 VanAvondt K, Nur E, Zeerleder S: Mechanisms of haemolysis-induced kidney injury. *Nat Rev Nephrol* 2019;15:671-692.
- 65 Brugnara C, Gee B, Armsby CC, Kurth S, Sakamoto M, Rifai N, et al.: Therapy with oral clotrimazole induces inhibition of the Gardos channel and reduction of erythrocyte dehydration in patients with sickle cell disease. *J Clin Invest* 1996;97:1227-1234.
- 66 Andolfo I, Russo R, Manna F, Shmukler BE, Gambale A, Vitiello G, et al.: Novel Gardos channel mutations linked to dehydrated hereditary stomatocytosis (xerocytosis). *Am J Hematol* 2015;90:921-926.
- 67 Rapetti-Mauss R, Lacoste C, Picard V, Guitton C, Lombard E, Loosveld M, et al.: A mutation in the Gardos channel is associated with hereditary xerocytosis. *Blood* 2015;126:1273-1280.
- 68 McGoron AJ, Joiner CH, Palascak MB, Claussen WJ, Franco RS: Dehydration of mature and immature sickle red blood cells during fast oxygenation/deoxygenation cycles: Role of KCl cotransport and extracellular calcium. *Blood* 2000;95:2164-2168.
- 69 Sebastiani P, Solovieff N, Hartley SW, Milton JN, Riva A, Dworkis DA, et al.: Genetic modifiers of the severity of sickle cell anemia identified through a genome-wide association study. *Am J Hematol* 2010;85:29-35.
- 70 Johnson RM, Gannon SA: Erythrocyte cation permeability induced by mechanical stress: A model for sickle cell cation loss. *Am J Physiol Cell Physiol* 1990;259:C746-751.
- 71 DeFranceschi L, Beuzard Y, Jouault H, Brugnara C: Modulation of erythrocyte potassium chloride cotransport, potassium content, and density by dietary magnesium intake in transgenic SAD mouse. *Blood* 1996;88:2738-2744.
- 72 Brugnara C, Chambers LA, Malynn E, Goldberg MA, Kruskall MS: Red blood cell regeneration induced by subcutaneous recombinant erythropoietin: Iron-deficient erythropoiesis in iron-replete subjects. *Blood* 1993;81:956-964.
- 73 Stocker JW, DeFranceschi L, McNaughton-Smith GA, Corrocher R, Beuzard Y, Brugnara C: ICA-17043, a novel Gardos channel blocker, prevents sickled red blood cell dehydration in vitro and in vivo in SAD mice. *Blood* 2003;101:2412-2418.
- 74 Gregg D, Goldschmidt-Clermont PJ: Cardiology patient page. Platelets and cardiovascular disease. *Circulation* 2003;108:1-3.
- 75 Bonnet D, Martin D, DeLonlay P, Villain E, Jouvet P, Rabier D, et al.: Arrhythmias and conduction defects as presenting symptoms of fatty acid oxidation disorders in children. *Circulation* 1999;100:2248-2253.
- 76 Lee VH, Thakur G, Nimjee SM, Youssef PP, Lakhani S, Heaton S, et al.: Early neurologic decline in acute ischemic stroke patients receiving thrombolysis with large vessel occlusion and mild deficits. *J Neurointerv Surg* 2020;1-3.
- 77 Thygesen K, Alpert JS, White HD: Universal Definition of Myocardial Infarction. *J Am Coll Cardiol* 2007;50:2173-2195.
- 78 Fernandes VRS, Polak JF, Edvardsen T, Carvalho B, Gomes A, Bluemke DA, et al.: Subclinical Atherosclerosis and Incipient Regional Myocardial Dysfunction in Asymptomatic Individuals. The Multi-Ethnic Study of Atherosclerosis (MESA). *J Am Coll Cardiol* 2006;47:2420-2428.
- 79 Brugada P, Abdollah H, Wellens HJJ: Continuous electrical activity during sustained monomorphic ventricular tachycardia. Observations on its dynamic behavior during the arrhythmia. *Am J Cardiol* 1985;55:402-411.
- 80 Tabuchi A, Hirai G, Saito J, Katsube Y, Sato H: Some problems in radiation exposure of pregnant women. *Hiroshima J Med Sci* 1975;24:213-228.
- 81 DelValle-Rodríguez A, López-Barneo J, Ureña J: Ca²⁺ channel-sarcoplasmic reticulum coupling: A mechanism of arterial myocyte contraction without Ca²⁺ influx. *EMBO J* 2003;22:4337-4345.
- 82 Gurney AM, Osipenko ON, MacMillan D, McFarlane KM, Tate RJ, Kempson FEJ: Two-Pore Domain K Channel, TASK-1, in Pulmonary Artery Smooth Muscle Cells. *Circ Res* 2003;93:957-964.
- 83 Moudgil R, Michelakis ED, Archer SL: The role of k⁺ channels in determining pulmonary vascular tone, oxygen sensing, cell proliferation, and apoptosis: implications in hypoxic pulmonary vasoconstriction and pulmonary arterial hypertension. *Microcirculation* 2006;13:615-632.

- 84 Domingues-Montanari S, Fernández-Cadenas I, delRío-Espinola A, Mendioroz M, Fernandez-Morales J, Corbeto N, et al.: KCNK17 genetic variants in ischemic stroke. *Atherosclerosis* 2010;208:203–209.
- 85 Ma Q, Wang Y, Shen Y, Liu X, Zhu X, Zhang H, et al.: The rs10947803 SNP of KCNK17 is associated with cerebral hemorrhage but not ischemic stroke in a Chinese population. *Neurosci Lett* 2013;539:82–85.
- 86 Roberts R: A genetic basis for coronary artery disease. *Trends Cardiovasc Med* 2015;25:171–178.
- 87 Winsvold BS, Bettella F, Witoelar A, Anttila V, Gormley P, Kurth T, et al.: Shared genetic risk between migraine and coronary artery disease: A genome-wide analysis of common variants. *PLoS One* 2017;12:1–15.
- 88 Christiansen MK, Larsen SB, Nyegaard M, Neergaard-Petersen S, Würtz M, Grove EL, et al.: The SH2B3 and KCNK5 loci may be implicated in regulation of platelet count, volume, and maturity. *Thromb Res* 2017;158:86–92.
- 89 Gestreau C, Heitzmann D, Thomas J, Dubreuil V, Bandulik S, Reichold M, et al.: Task2 potassium channels set central respiratory CO₂ and O₂ sensitivity. *Proc Natl Acad Sci U S A* 2010;107:2325–2330.
- 90 Staudacher K, Staudacher I, Ficker E, Seyler C, Gierten J, Kisselbach J, et al.: Carvedilol targets human K_{2P3.1} (TASK1) K⁺ leak channels. *Br J Pharmacol* 2011;163:1099–1110.
- 91 Wiedmann F, Kiper AK, Bedoya M, Ratte A, Rinné S, Kraft M, et al.: Identification of the A293 (AVE1231) binding site in the cardiac two-pore-domain potassium channel TASK-1: A common low affinity antiarrhythmic drug binding site. *Cell Physiol Biochem* 2019;52:1223–1235.
- 92 Gierten J, Ficker E, Bloehs R, Schweizer PA, Zitron E, Scholz E, et al.: The human cardiac K_{2P3.1} (TASK-1) potassium leak channel is a molecular target for the class III antiarrhythmic drug amiodarone. *Naunyn Schmiedebergs Arch Pharmacol* 2010;381:261–270.
- 93 Seyler C, Li J, Schweizer PA, Katus HA, Thomas D: Inhibition of cardiac two-pore-domain K⁺ (K_{2P}) channels by the antiarrhythmic drug vernakalant - Comparison with flecainide. *Eur J Pharmacol* 2014;724:51–57.
- 94 Navas P, Tenorio J, Quezada CA, Barrios E, Gordo G, Arias P, et al.: Molecular Analysis of BMPR2, TBX4, and KCNK3 and Genotype-Phenotype Correlations in Spanish Patients and Families With Idiopathic and Hereditary Pulmonary Arterial Hypertension. *Rev Esp Cardiol (Engl Ed)* 2016;69:1011–1019.
- 95 Cunningham KP, Holden RG, Escribano-Subias PM, Cogolludo A, Veale EL, Mathie A: Characterization and regulation of wild-type and mutant TASK-1 two pore domain potassium channels indicated in pulmonary arterial hypertension. *J Physiol* 2019;597:1087–1101.
- 96 Ma L, Roman-Campos D, Austin ED, Eyries M, Sampson KS, Soubrier F, et al.: A novel channelopathy in pulmonary arterial hypertension. *N Engl J Med* 2013;369:351–361.
- 97 Schmidt C, Wiedmann F, Schweizer PA, Katus HA, Thomas D: Inhibition of cardiac two-pore-domain K⁺ (K_{2P}) channels - An emerging antiarrhythmic concept. *Eur J Pharmacol* 2014;738:250–255.
- 98 Lugenbiel P, Wenz F, Syren P, Geschwill P, Govorov K, Seyler C, et al.: TREK-1 (K_{2P2.1}) K⁺ channels are suppressed in patients with atrial fibrillation and heart failure and provide therapeutic targets for rhythm control. *Basic Res Cardiol* 2017;112:1–14.
- 99 Decher N, Ortiz-Bonnin B, Friedrich C, Schewe M, Kiper AK, Rinné S, et al.: Sodium permeable and “hypersensitive” TREK -1 channels cause ventricular tachycardia. *EMBO Mol Med* 2017;9:403–414.
- 100 Friedrich C, Rinné S, Zumhagen S, Kiper AK, Silbernage IN, Netter MF, et al.: Gain-of-function mutation in TASK -4 channels and severe cardiac conduction disorder. *EMBO Mol Med* 2014;6:937–951.
- 101 Chai S, Wan X, Ramirez-Navarro A, Tesar PJ, Kaufman ES, Ficker E, et al.: Physiological genomics identifies genetic modifiers of long QT syndrome type 2 severity. *J Clin Invest* 2018;128:1043–1056.
- 102 Staudacher I, Illg C, Chai S, Deschenes I, Seehausen S, Gramlich D, et al.: Cardiovascular pharmacology of K_{2P 17.1} (TASK-4, TALK-2) two-pore-domain K⁺ channels. *Naunyn Schmiedebergs Arch Pharmacol* 2018;391:1119–1131.
- 103 Staudacher I, Illg C, Gierten J, Seehausen S, Schweizer PA, Katus HA, et al.: Identification and functional characterization of zebrafish K(2P)17.1 (TASK-4, TALK-2) two-pore-domain K(+) channels. *Eur J Pharmacol* 2018;831:94–102.
- 104 Yang X, Guo P, Li J, Wang W, Xu S, Wang L, et al.: Functional study of TREK-1 potassium channels during rat heart development and cardiac ischemia using RNAi techniques. *J Cardiovasc Pharmacol* 2014;64:142–150.
- 105 Meuth SG, Kleinschnitz C, Broicher T, Austinat M, Braeuninger S, Bittner S, et al.: The neuroprotective impact of the leak potassium channel TASK1 on stroke development in mice. *Neurobiol Dis* 2009;33:1–11.
- 106 Laigle C, Confort-Gouny S, LeFur Y, Cozzone PJ, Viola A: Deletion of TRAAK Potassium Channel Affects Brain Metabolism and Protects against Ischemia. *PLoS One* 2012;7:1–12.

- 107 Aimond F, Rauzier JM, Bony C, Vassort G: Simultaneous activation of p38 MAPK and p42/44 MAPK by ATP stimulates the K⁺ current ITREK in cardiomyocytes. *J Biol Chem* 2000;275:39110–39116.
- 108 Tang B, Li Y, Nagaraj C, Morty RE, Gabor S, Stacher E, et al.: Endothelin-1 inhibits background two-pore domain channel TASK-1 in primary human pulmonary artery smooth muscle cells. *Am J Respir Cell Mol Biol* 2009;41:476–483.
- 109 Hund TJ, Snyder JS, Wu X, Glynn P, Koval OM, Onal B, et al.: β iV-Spectrin regulates TREK-1 membrane targeting in the heart. *Cardiovasc Res* 2014;102:166–175.
- 110 Unudurthi SD, Wu X, Qian L, Amari F, Onal B, Li N, et al.: Two-Pore K⁺ channel TREK-1 regulates sinoatrial node membrane excitability. *J Am Heart Assoc* 2016;5:e002865.
- 111 Lugenbiel P, Govorov K, Rahm AK, Wieder T, Gramlich D, Syren P, et al.: Inhibition of Histone Deacetylases Induces K⁺ Channel Remodeling and Action Potential Prolongation in HL-1 Atrial Cardiomyocytes. *Cell Physiol Biochem* 2018;49:65–77.
- 112 Abraham DM, Wolf MJ, Howard A, Invest JC, Abraham DM, Lee TE, et al.: The two-pore domain potassium channel TREK-1 mediates cardiac fibrosis and diastolic dysfunction Graphical abstract Find the latest version: The two-pore domain potassium channel TREK-1 mediates cardiac fibrosis and diastolic dysfunction. *J Clin Invest* 2018;128:4843–4855.
- 113 Wiedmann F, Schulte JS, Gomes B, Zafeiriou MP, Ratte A, Rathjens F, et al.: Atrial fibrillation and heart failure-associated remodeling of two-pore-domain potassium (K_{2p}) channels in murine disease models: focus on TASK-1. *Basic Res Cardiol* 2018;113:1–14.
- 114 Donner BC, Schullenberg M, Geduldig N, Hüning A, Mersmann J, Zacharowski K, et al.: Functional role of TASK-1 in the heart: Studies in TASK-1-deficient mice show prolonged cardiac repolarization and reduced heart rate variability. *Basic Res Cardiol* 2011;106:75–87.
- 115 Petric S, Clasen L, vanWessel C, Geduldig N, Ding Z, Schullenberg M, et al.: In vivo electrophysiological characterization of TASK-1 deficient mice. *Cell Physiol Biochem Int J Exp Cell Physiol Biochem Pharmacol* 2012;30:523–537.
- 116 Bauer CK, Calligari P, Radio FC, Caputo V, Dentici ML, Falah N, et al.: Mutations in KCNK4 that Affect Gating Cause a Recognizable Neurodevelopmental Syndrome. *Am J Hum Genet* 2018;103:621–630.
- 117 Graham JM, Zadeh N, Kelley M, Tan ES, Liew W, Tan V, et al.: KCNK9 imprinting syndrome—further delineation of a possible treatable disorder. *Am J Med Genet Part A* 2016;170:2632–2637.
- 118 Li J, Loebel A, Meltzer HY: Identifying the genetic risk factors for treatment response to lurasidone by genome-wide association study: A meta-analysis of samples from three independent clinical trials. *Schizophr Res* 2018;199:203–213.
- 119 Rukova B, Staneva R, Hadjidekova S, Stamenov G, Milanova V, Toncheva D: Whole genome methylation analyses of schizophrenia patients before and after treatment. *Biotechnol Biotechnol Equip* 2014;28:518–524.
- 120 Liu J, Siyahhan Julnes P, Chen J, Ehrlich S, Walton E, Calhoun VD: The association of DNA methylation and brain volume in healthy individuals and schizophrenia patients. *Schizophr Res* 2015;169:447–452.
- 121 Perlis RH, Moorjani P, Fagerness J, Purcell S, Trivedi MH, Fava M, et al.: Pharmacogenetic analysis of genes implicated in rodent models of antidepressant response: Association of TREK1 and treatment resistance in the STAR*D study. *Neuropsychopharmacology* 2008;33:2810–2819.
- 122 Rainero I, Rubino E, Gallone S, Zavarise P, Carli D, Boschi S, et al.: KCNK18 (TRESK) genetic variants in Italian patients with migraine. *Headache* 2014;54:1515–1522.
- 123 Andres-Enguix I, Shang L, Stansfeld PJ, Morahan JM, Sansom MSP, Lafrenière RG, et al.: Functional analysis of missense variants in the TRESK (KCNK18) K⁺ channel. *Sci Rep* 2012;2:1–7.
- 124 Maher BH, Taylor M, Stuart S, Okolicsanyi RK, Roy B, Sutherland HG, et al.: Analysis of 3 common polymorphisms in the KCNK18 gene in an Australian Migraine Case-control cohort. *Gene* 2013;528:343–346.
- 125 Pettingill P, Weir GA, Wei T, Wu Y, Flower G, Lalic T, et al.: A causal role for TRESK loss of function in migraine mechanisms. *Brain* 2019;142:3852–3867.
- 126 Veale EL, Mathie A: Aristolochic acid, a plant extract used in the treatment of pain and linked to Balkan endemic nephropathy, is a regulator of K_{2p} channels. *Br J Pharmacol* 2016;173:1639–1652.
- 127 Gormley P, Winsvold BS, Nyholt DR, Kallela M, Chasman DI, Palotie A: Migraine genetics: From genome-wide association studies to translational insights. *Genome Med* 2016;8:8–10.

- 128 Sáez-Hernández L, Peral B, Sanz R, Gómez-Garre P, Ramos C, Ayuso C, et al.: Characterization of a 6p21 translocation breakpoint in a family with idiopathic generalized epilepsy. *Epilepsy Res* 2003;56:155–163.
- 129 Xiao Z, Deng PY, Rojanathammanee L, Yang C, Grisanti L, Permpoonputtana K, et al.: Noradrenergic depression of neuronal excitability in the entorhinal cortex activation of TREK-2 K⁺ channels. *J Biol Chem* 2009;284:10980–10991.
- 130 Meadows HJ, Chapman CG, Duckworth DM, Kellsell RE, Murdock PR, Nasir S, et al.: The neuroprotective agent sipatrigine (BW619C89) potentially inhibits the human tandem pore-domain K⁺ channels TREK-1 and TRAAK. *Brain Res* 2001;892:94–101.
- 131 Xi G, Zhang X, Zhang L, Sui Y, Hui J, Liu S, et al.: Fluoxetine attenuates the inhibitory effect of glucocorticoid hormones on neurogenesis in vitro via a two-pore domain potassium channel, TREK-1. *Psychopharmacology (Berl)* 2011;214:747–759.
- 132 Heurteaux C, Lucas G, Guy N, ElYacoubi M, Thümmeler S, Peng XD, et al.: Deletion of the background potassium channel TREK-1 results in a depression-resistant phenotype. *Nat Neurosci* 2006;9:1134–1141.
- 133 Borsotto M, Veyssiere J, Moha Ou Maati H, Devader C, Mazella J, Heurteaux C: Targeting two-pore domain K⁺ channels TREK-1 and TASK-3 for the treatment of depression: A new therapeutic concept. *Br J Pharmacol* 2015;172:771–784.
- 134 Mazella J, Pétrault O, Lucas G, Deval E, Béraud-Dufour S, Gandin C, et al.: Spadin, a sortilin-derived peptide, targeting rodent TREK-1 channels: A new concept in the antidepressant drug design. *PLoS Biol* 2010;8:e1000355.
- 135 Ye D, Li Y, Zhang X, Guo F, Geng L, Zhang Q, et al.: TREK1 channel blockade induces an antidepressant-like response synergizing with 5-HT_{1A} receptor signaling. *Eur Neuropsychopharmacol* 2015;25:2426–2436.
- 136 Joseph A, Thuy TTT, Thanh LT, Okada M: Antidepressive and anxiolytic effects of ostruthin, a TREK-1 channel activator. *PLoS One* 2018;13:1–19.
- 137 Alloui A, Zimmermann K, Mamet J, Duprat F, Noël J, Chemin J, et al.: TREK-1, a K⁺ channel involved in polymodal pain perception. *EMBO J* 2006;25:2368–2376.
- 138 Devilliers M, Busserolles J, Lolignier S, Deval E, Pereira V, Alloui A, et al.: Activation of TREK-1 by morphine results in analgesia without adverse side effects. *Nat Commun* 2013;4:2941.
- 139 Dadi PK, Vierra NC, Days E, Dickerson MT, Vinson PN, Weaver CD, et al.: Selective Small Molecule Activators of TREK-2 Channels Stimulate Dorsal Root Ganglion c-Fiber Nociceptor Two-Pore-Domain Potassium Channel Currents and Limit Calcium Influx. *ACS Chem Neurosci* 2017;8:558–568.
- 140 Acosta C, Djouhri L, Watkins R, Berry C, Bromage K, Lawson SN: TREK2 expressed selectively in IB4-binding C-fiber nociceptors hyperpolarizes their membrane potentials and limits spontaneous pain. *J Neurosci* 2014;34:1494–1509.
- 141 Liu P, Xiao Z, Ren F, Guo Z, Chen Z, Zhao H, et al.: Functional analysis of a migraine-associated TRESK K⁺ channel mutation. *J Neurosci* 2013;33:12810–12824.
- 142 Guo Z, Cao YQ: Over-expression of TRESK K⁺ channels reduces the excitability of trigeminal ganglion nociceptors. *PLoS One* 2014;9:e87029.
- 143 Helderman JH, Elahi D, Andersen DK, Raizes GS, Tobin JD, Shocken D, et al.: Prevention of the glucose intolerance of thiazide diuretics by maintenance of body potassium. *Diabetes* 1983;32:106–111.
- 144 Rowe JW, Tobin JD, Rosa RM, Andres R: Effect of experimental potassium deficiency on glucose and insulin metabolism. *Metabolism* 1980;29:498–502.
- 145 Colditz GA, Manson JE, Stampfer MJ, Rosner B, Willett WC, Speizer FE: Diet and risk of clinical diabetes in women. *Am J Clin Nutr* 1992;55:1018–1023.
- 146 Chatterjee R, Colangelo LA, Yeh HC, Anderson CA, Daviglius ML, Liu K, et al.: Potassium intake and risk of incident type 2 diabetes mellitus: The Coronary Artery Risk Development in Young Adults (CARDIA) Study. *Diabetologia* 2012;55:1295–1303.
- 147 Cho YS, Chen C, Hu C, Long J, Hee RT, Sim X, et al.: NIH Public Access new loci for type 2 diabetes in East Asians. *Nat Genet* 2013;44:67–72.
- 148 Wood AR, Jonsson A, Jackson AU, Wang N, VanLeewen N, Palmer ND, et al.: A genome-wide association study of IVGTT-based measures of first-phase insulin secretion refines the underlying physiology of type 2 diabetes variants. *Diabetes* 2017;66:2296–2309.
- 149 van deBunt M, Gaulton KJ, Parts L, Moran I, Johnson PR, Lindgren CM, et al.: The miRNA Profile of Human Pancreatic Islets and Beta-Cells and Relationship to Type 2 Diabetes Pathogenesis. *PLoS One* 2013;8:1–7.

- 150 Graff SM, Johnson SR, Leo PJ, Dadi PK, Nakhe AY, McInerney-Leo AM, et al.: A novel mutation in KCNK16 causing a gain-of-function in the TALK-1 potassium channel: a new cause of maturity onset diabetes of the young. *bioRxiv* 2020;2020.02.04.929430.
- 151 Dadi PK, Vierra NC, Jacobson DA: Pancreatic β -cell-specific ablation of TASK-1 channels augments glucose-stimulated calcium entry and insulin secretion, improving glucose tolerance. *Endocrinology* 2014;155:3757–3768.
- 152 Vierra NC, Dadi PK, Jeong I, Dickerson M, Powell DR, Jacobson DA: Type 2 diabetes-associated K⁺ channel TALK-1 modulates β -cell electrical excitability, second-phase insulin secretion, and glucose homeostasis. *Diabetes* 2015;64:3818–3828.
- 153 Vierra NC, Dadi PK, Milian SC, Dickerson MT, Jordan KL, Gilon P, et al.: TALK-1 channels control β cell endoplasmic reticulum Ca²⁺ homeostasis. *Sci Signal* 2017;10:eaan2883.
- 154 Dickerson MT, Bogart AM, Altman MK, Milian SC, Jordan KL, Dadi PK, et al.: Cytokine-mediated changes in K⁺ channel activity promotes an adaptive Ca²⁺ response that sustains β -cell insulin secretion during inflammation. *Sci Rep* 2018;8:1–15.
- 155 Giebisch G: Renal potassium channels: Function, regulation, and structure. *Kidney Int* 2001;60:436–445.
- 156 Giebisch G: Renal potassium transport: Mechanisms and regulation. *Am J Physiol Ren Physiol* 1998;274:F817–833.
- 157 Ueda Y, Ookawara S, Ito K, Miyazawa H, Kaku Y, Hoshino T, et al.: Changes in urinary potassium excretion in patients with chronic kidney disease. *Kidney Res Clin Pract* 2016;35:78–83.
- 158 Mun EG, Park JE, Cha YS: Effects of doenjang, a traditional Korean soybean paste, with high-salt diet on blood pressure in Sprague–Dawley rats. *Nutrients* 2019;11:2745.
- 159 Reyes R, Duprat F, Lesage F, Fink M, Salinas M, Farman N, et al.: Cloning and expression of a novel pH-sensitive two pore domain K⁺ channel from human kidney. *J Biol Chem* 1998;273:30863–30869.
- 160 Pineda RH, Nedumaran B, Hypolite J, Pan XQ, Wilson S, Meacham RB, et al.: Altered expression and modulation of the two-pore-domain (K2p) mechanogated potassium channel TREK-1 in overactive human detrusor. *Am J Physiol Ren Physiol* 2017;313:F535–F546.
- 161 Nedumaran B, Pineda RH, Rudra P, Lee S, Malykhina AP: Association of genetic polymorphisms in the pore domains of mechano-gated TREK-1 channel with overactive lower urinary tract symptoms in humans. *Neurourol Urodyn* 2019;38:144–150.
- 162 Warth R, Barrière H, Meneton P, Bloch M, Thomas J, Tauc M, et al.: Proximal renal tubular acidosis in TASK2 K⁺ channel-deficient mice reveals a mechanism for stabilizing bicarbonate transport. *Proc Natl Acad Sci U S A* 2004;101:8215–8220.
- 163 Pineda RH, Hypolite J, Lee S, Carrasco A, Iguchi N, Meacham RB, et al.: Altered detrusor contractility and voiding patterns in mice lacking the mechanosensitive TREK-1 channel. *BMC Urol* 2019;19:1–15.
- 164 Baker S, Hatton W, Han J, Hennig G, Britton F, Koh S: Role of TREK-1 Potassium Channel in Bladder Overactivity After Partial Bladder Outlet Obstruction in Mouse. *J Urol* 2009;183:793–800.
- 165 Zhang J, Cao M, Chen Y, Wan Z, Wang H, Lin H, et al.: Increased expression of TREK-1 K⁺ channel in the dorsal root ganglion of rats with detrusor overactivity after partial bladder outlet obstruction. *Med Sci Monit* 2018;24:1064–1071.
- 166 Alcaïno C, Farrugia G, Beyder A: Mechanosensitive Piezo Channels in the Gastrointestinal. *Curr Top Membr* 2017;79:219–244.
- 167 Ma R, Seifi M, Papanikolaou M, Brown JF, Swinny JD, Lewis A: TREK-1 channel expression in smooth muscle as a target for regulating murine intestinal contractility: Therapeutic implications for motility disorders. *Front Physiol* 2018;9:1–12.
- 168 Tomuschat C, O'Donnell AM, Coyle D, Dreher N, Kelly D, Puri P: Altered expression of a two-pore domain (K2P) mechano-gated potassium channel TREK-1 in Hirschsprung's disease. *Pediatr Res* 2016;80:729–733.
- 169 O'Donnell AM, Nakamura H, Parekh B, Puri P: Decreased expression of TRAAK channels in Hirschsprung's disease: a possible cause of postoperative dysmotility. *Pediatr Surg Int* 2019;35:1431–1435.
- 170 Cho SY, Beckett EA, Baker SA, Han I, Park KJ, Monaghan K, et al.: A pH-sensitive potassium conductance (TASK) and its function in the murine gastrointestinal tract. *J Physiol* 2005;565:243–259.
- 171 Julio-Kalajzić F, Villanueva S, Burgos J, Ojeda M, Cid LP, Jentsch TJ, et al.: K^{2p} TASK-2 and KCNQ1–KCNE3 K⁺ channels are major players contributing to intestinal anion and fluid secretion. *J Physiol* 2018;596:393–407.

- 172 Currò D: The Modulation of Potassium Channels in the Smooth Muscle as a Therapeutic Strategy for Disorders of the Gastrointestinal Tract. *Adv Protein Chem Struct Biol* 2016;104:263-305.
- 173 Konakov MV, Berezhnov AV, Teplov IY, Levin SG, Godukhin OV: Identification and properties of bupivacaine-sensitive potassium currents in cultured hippocampal neurons. *Biochem Moscow Suppl Ser A* 2015;9:309-317.
- 174 Shin HW, Soh JS, Kim HZ, Hong J, Woo DH, Heo JY, et al.: The inhibitory effects of bupivacaine, levobupivacaine, and ropivacaine on K2P (two-pore domain potassium) channel TREK-1. *J Anesth* 2014;28:81-86.
- 175 Barekattain A, Razavi M: Antiarrhythmic therapy in atrial fibrillation: Indications, guidelines, and safety. *Texas Hear Inst J* 2012;39:532-534.
- 176 Bichet D, Blin S, Feliciangeli S, Chatelain FC, Bobak N, Lesage F: Silent but not dumb: How cellular trafficking and pore gating modulate expression of TWIK1 and THIK2. *Pflugers Arch Eur J Physiol* 2015;467:1121-1131.
- 177 Bobak N, Feliciangeli S, Chen CC, Soussia IB, Bittner S, Pagnotta S, et al.: Recombinant tandem of pore-domains in a Weakly Inward rectifying K⁺ channel 2 (TWIK2) forms active lysosomal channels. *Sci Rep* 2017;7:1-13.
- 178 Pandit LM, Lloyd EE, Reynolds JO, Lawrence WS, Reynolds C, Wehrens XHT, et al.: TWIK-2 channel deficiency leads to pulmonary hypertension through a rho-kinase-mediated process. *Hypertension* 2014;64:1260-1265.
- 179 Kitagawa MG, Reynolds JO, Durgan D, Rodney G, Karmouty-Quintana H, Bryan R, et al.: Twik-2 ^{-/-} mouse demonstrates pulmonary vascular heterogeneity in intracellular pathways for vasocontractility. *Physiol Rep* 2019;7:1-9.
- 180 Di A, Xiong S, Ye Z, Malireddi RKS, Kometani S, Zhong M, et al.: The TWIK2 Potassium Efflux Channel in Macrophages Mediates NLRP3 Inflammasome-Induced Inflammation. *Immunity* 2018;49:56-65.e4.